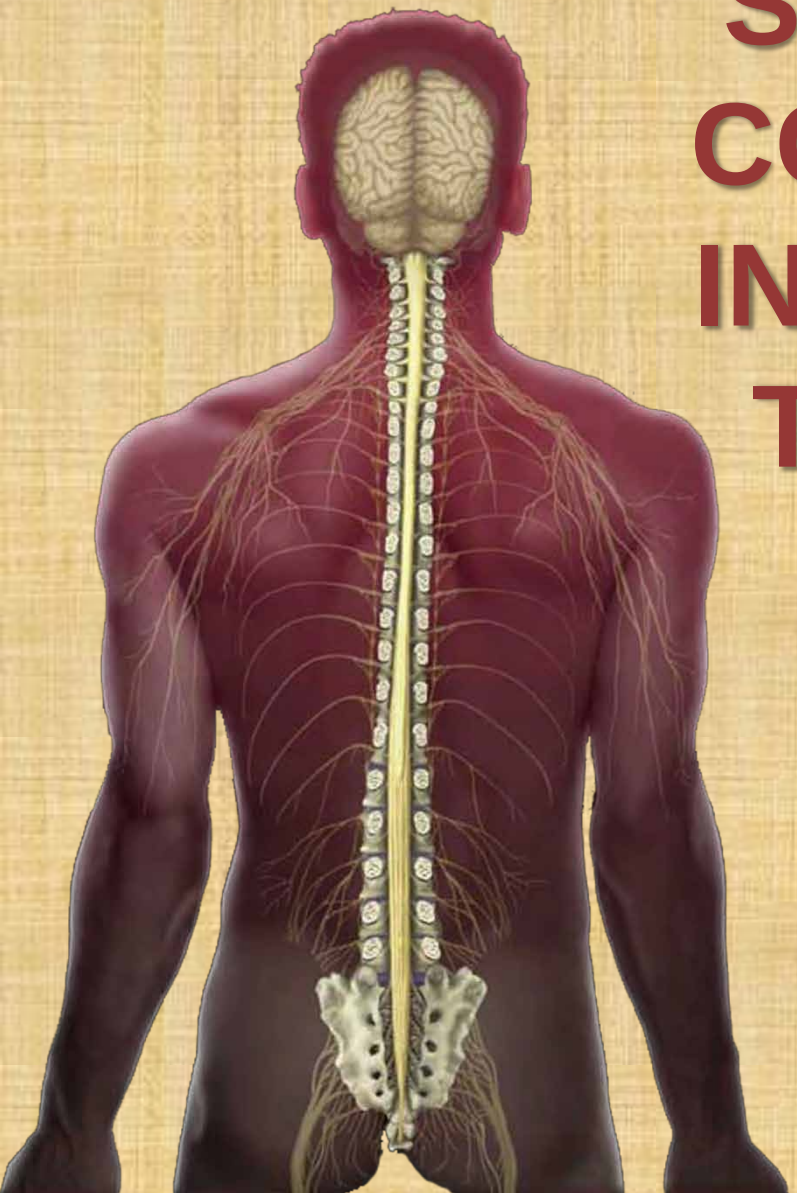
CENTRAL NERVOUS SYSTEM AND THE CONTRIBUTION OF INFLAMMATION TO TISSUE DAMAGE AFTER INJURY



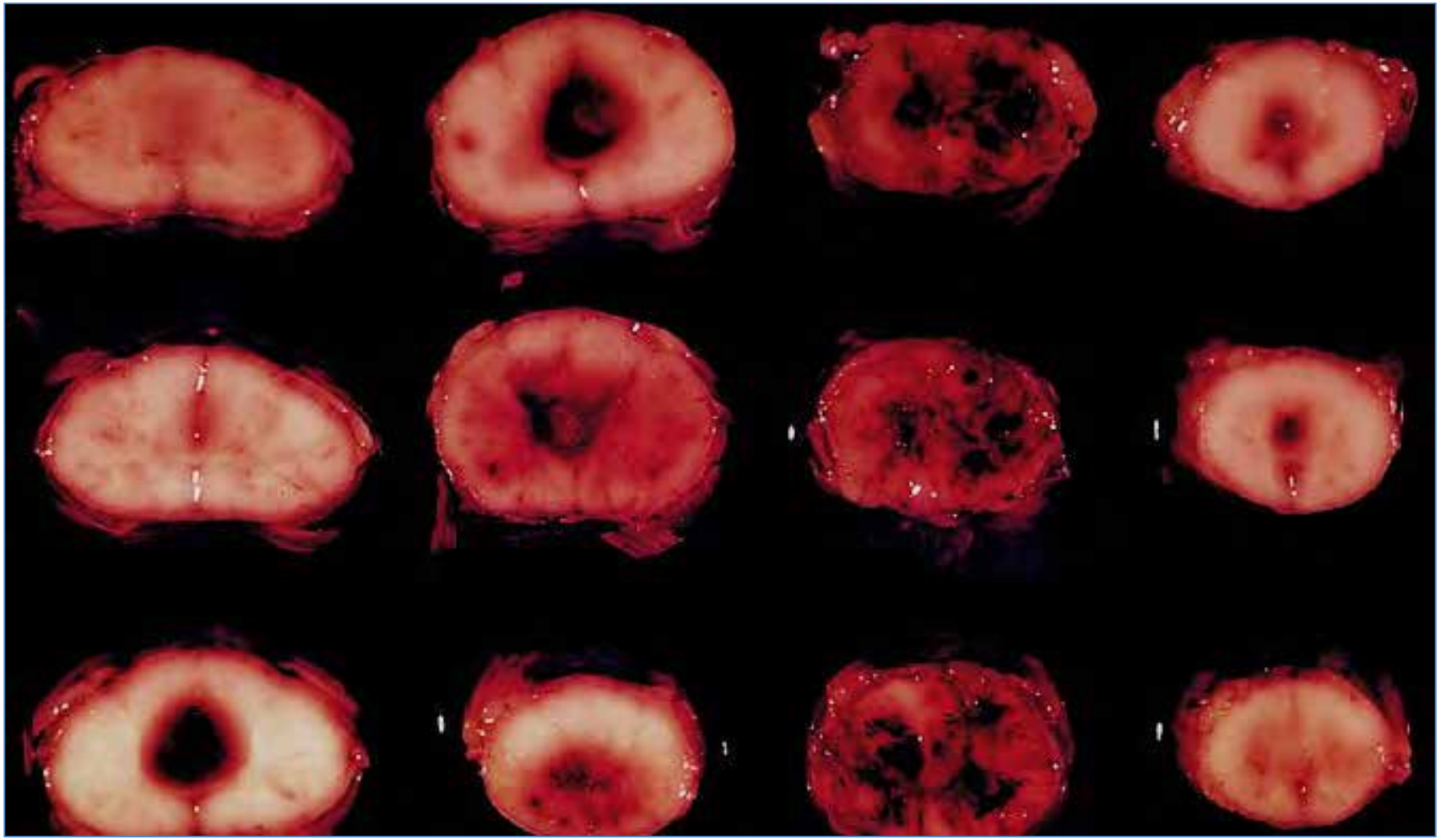
Rubèn López-Vales PhD

Institut de Neurociències

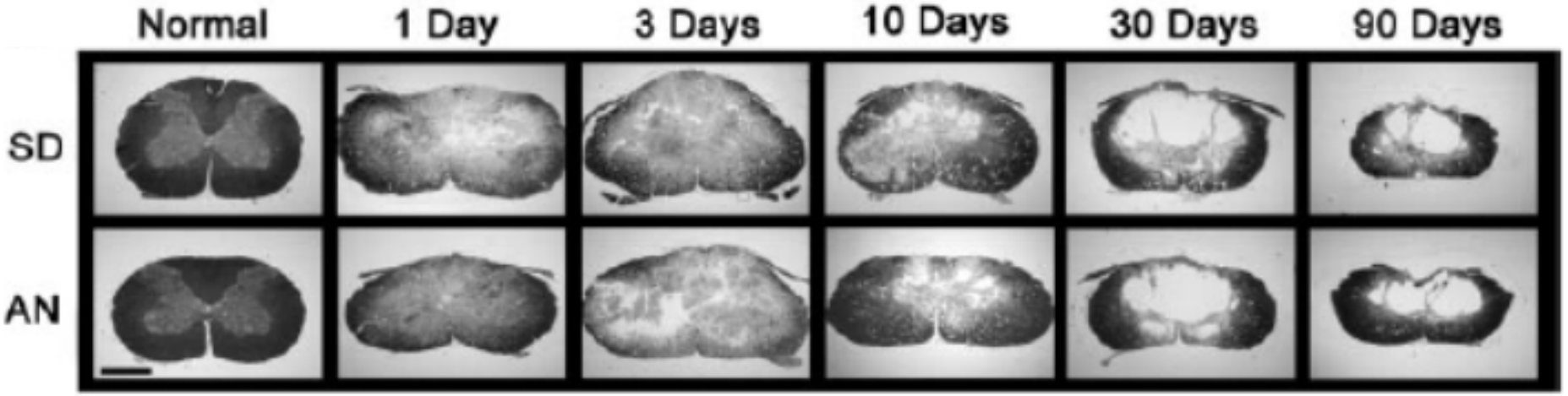
Department of Cell Biology, Physiology and Immunology

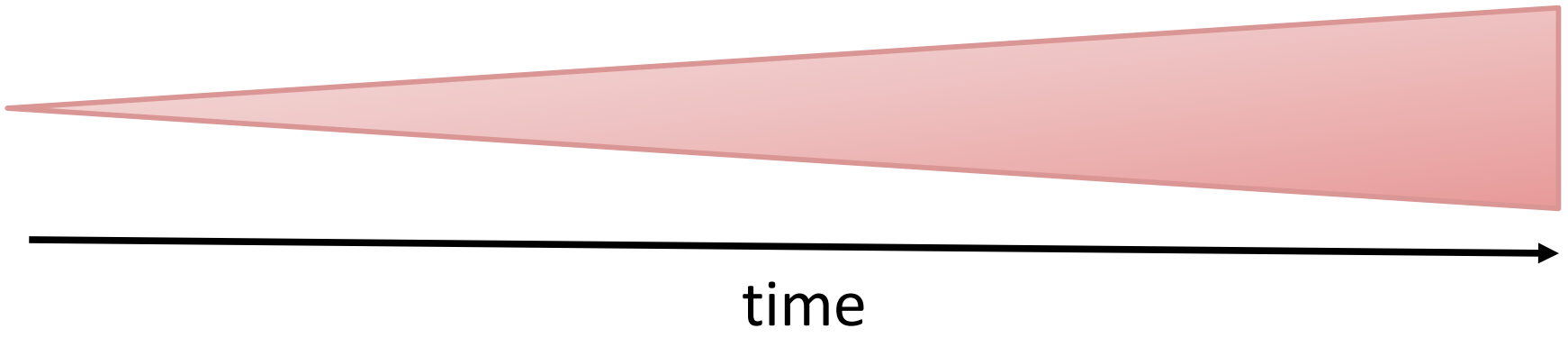
Universitat Autònoma de Barcelona

Spinal cord injury: tissue degeneration

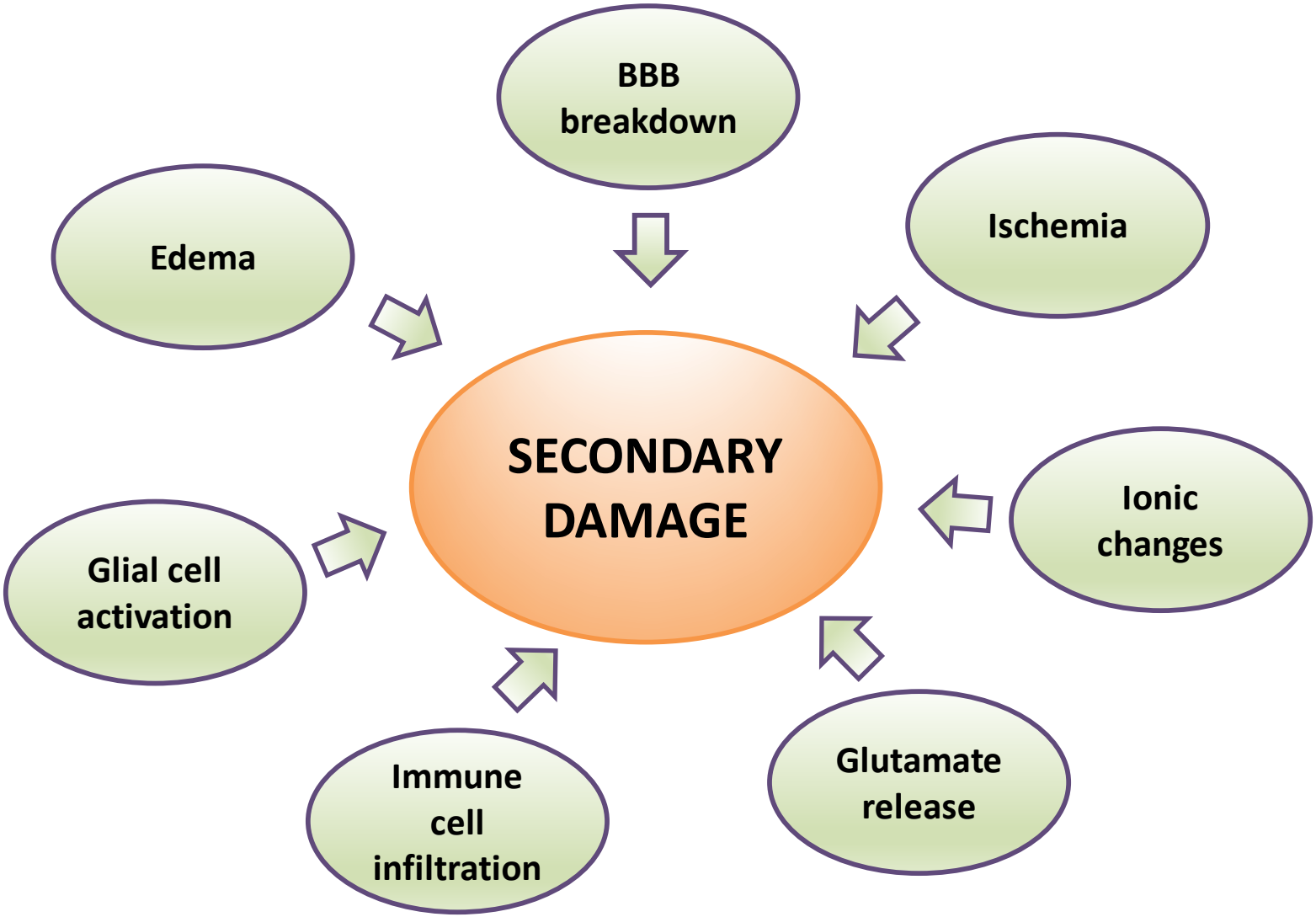


Spinal cord injury: secondary degeneration

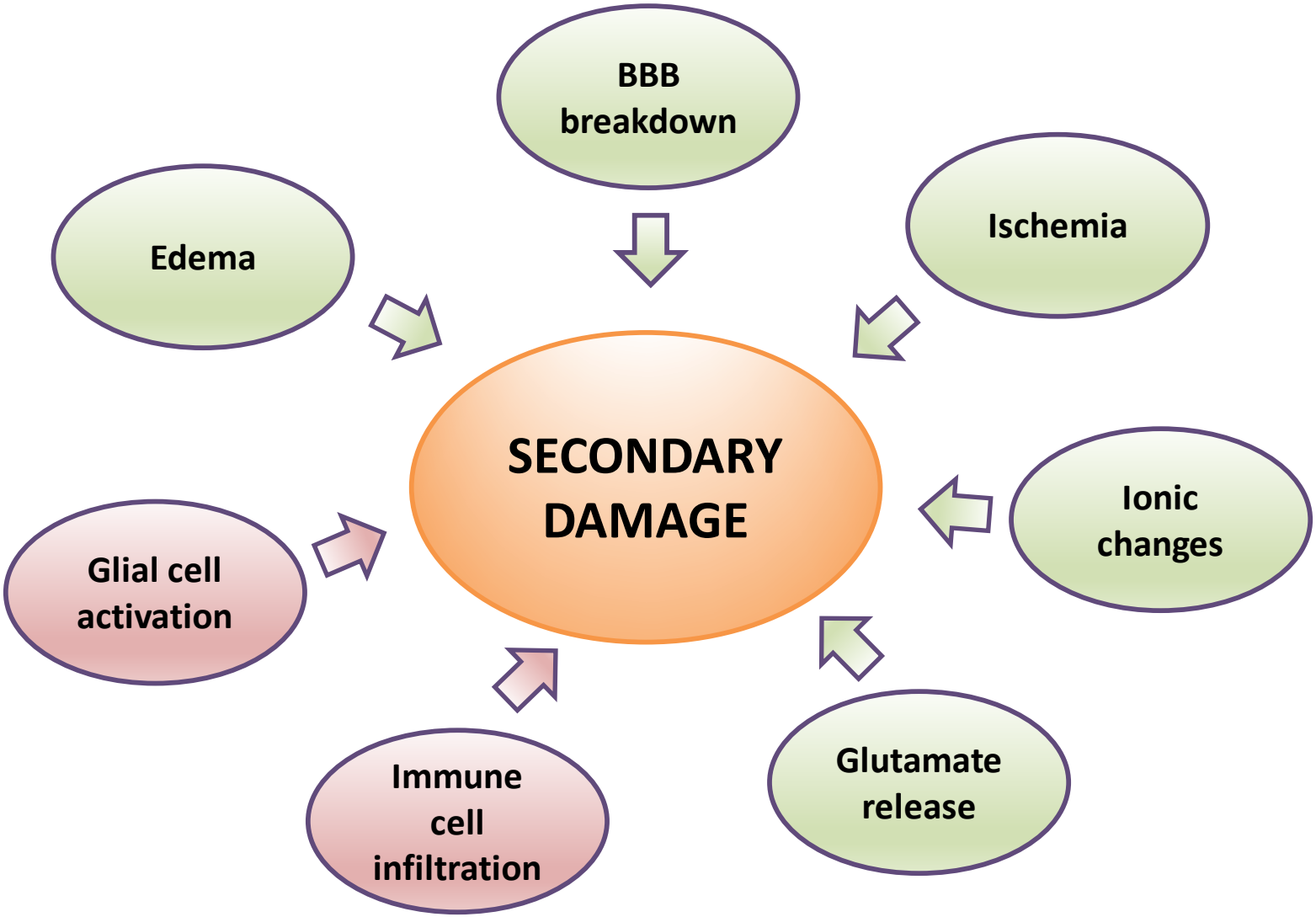


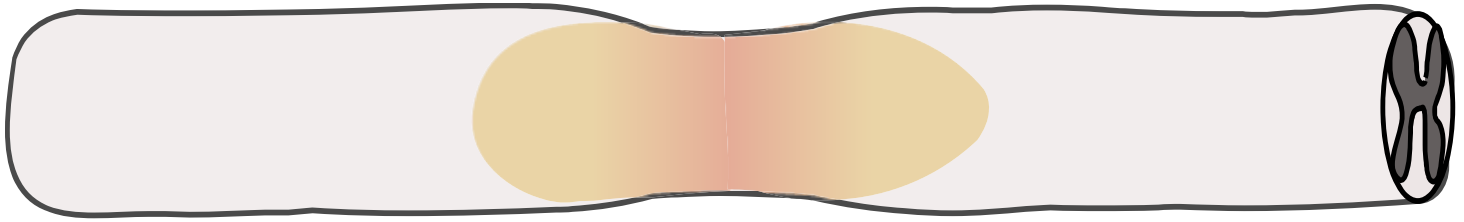


Spinal cord injury: secondary degeneration

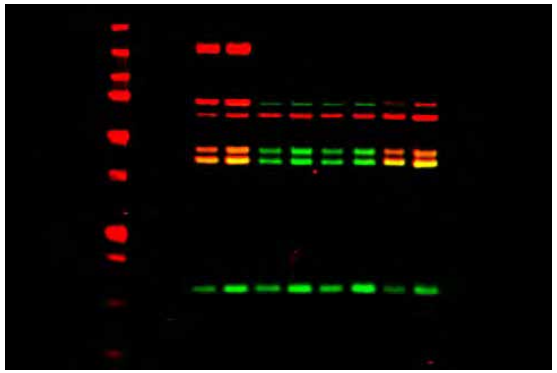


Spinal cord injury: secondary degeneration



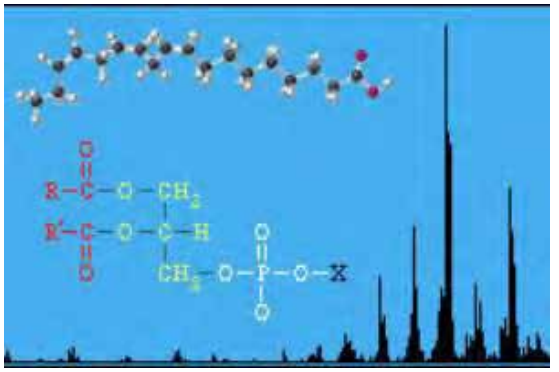


Proteins

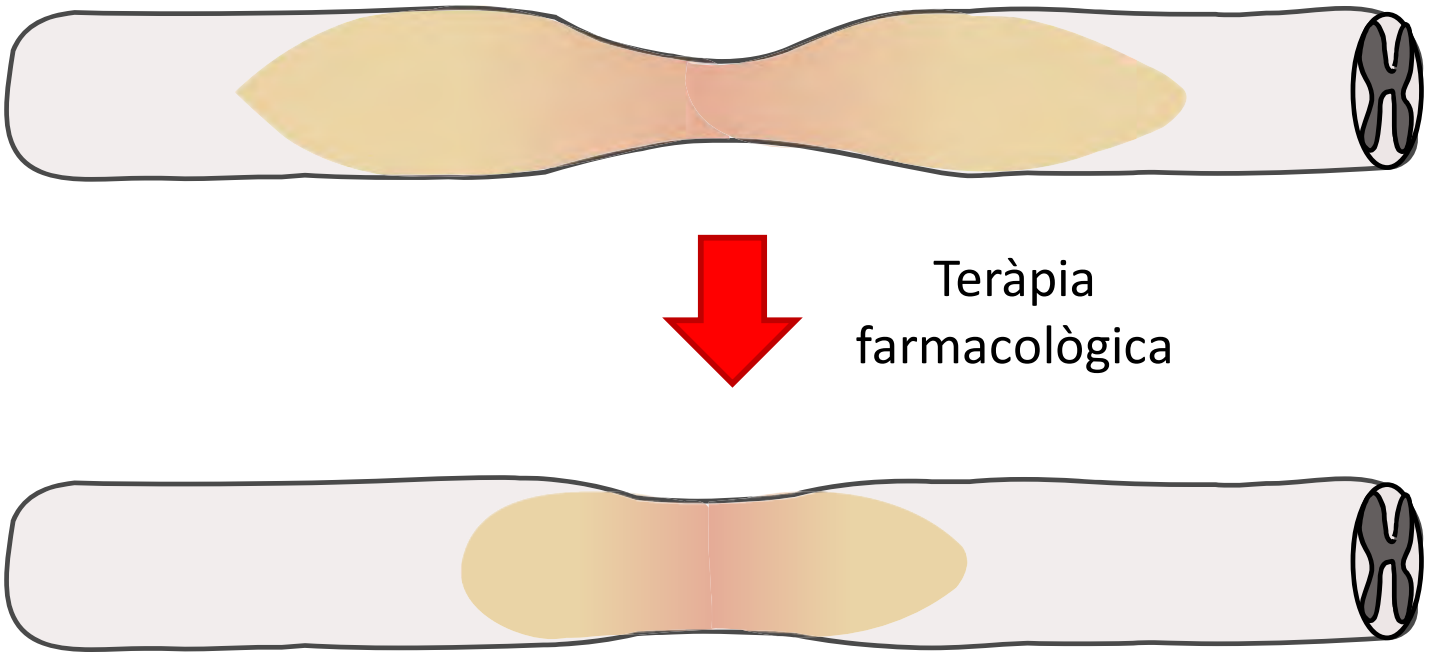


ARN

Lipids

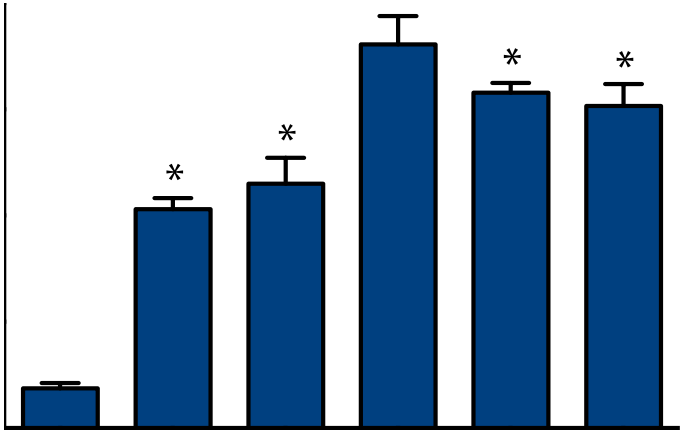


THERAPEUTIC APPROACHES


Teràpia
farmacològica

Lysophosphatidic Acid

LPA levels increase in the spinal cord tissue after injury.

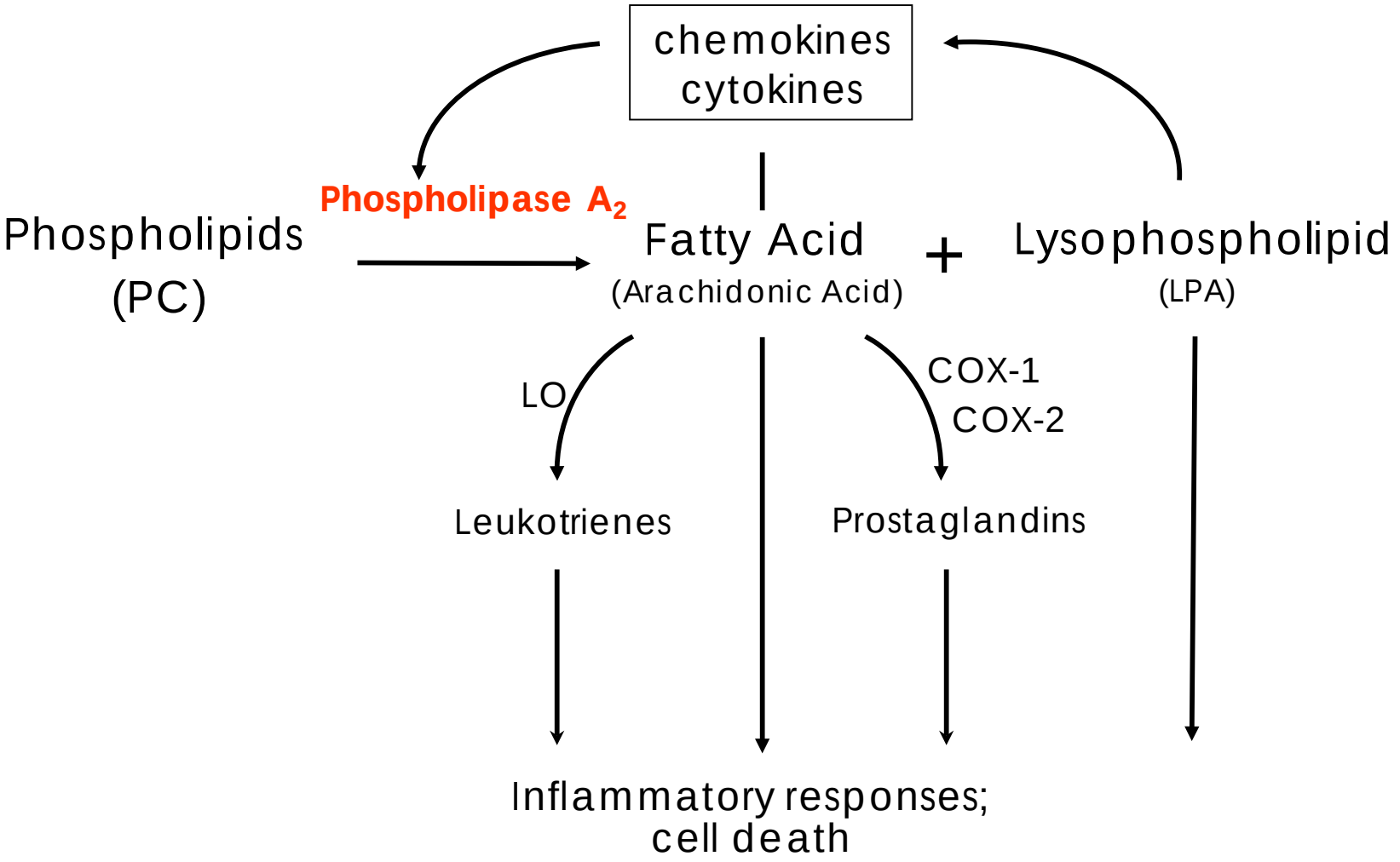


* p < 0,05 vs. naïve

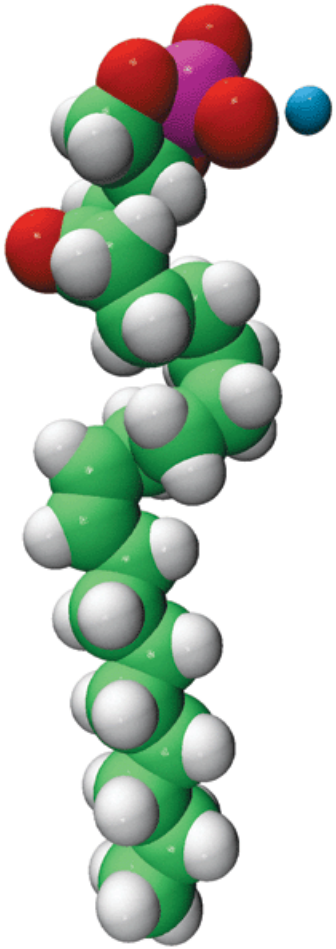
	16:0	18:1	18:0	20:4	20:1	22:6	22:4	22:1
Naïve SC	✗	✓	✓	✗	✓	✗	✗	✗
Injured SC	✓	✓	✓	✓	✓	✓	✓	✓

Santos-Nogueira et al., 2015 J Neurosci

Lysophosphatidic Acid



Lysophosphatidic Acid



LPA has wide variety of biological functions:

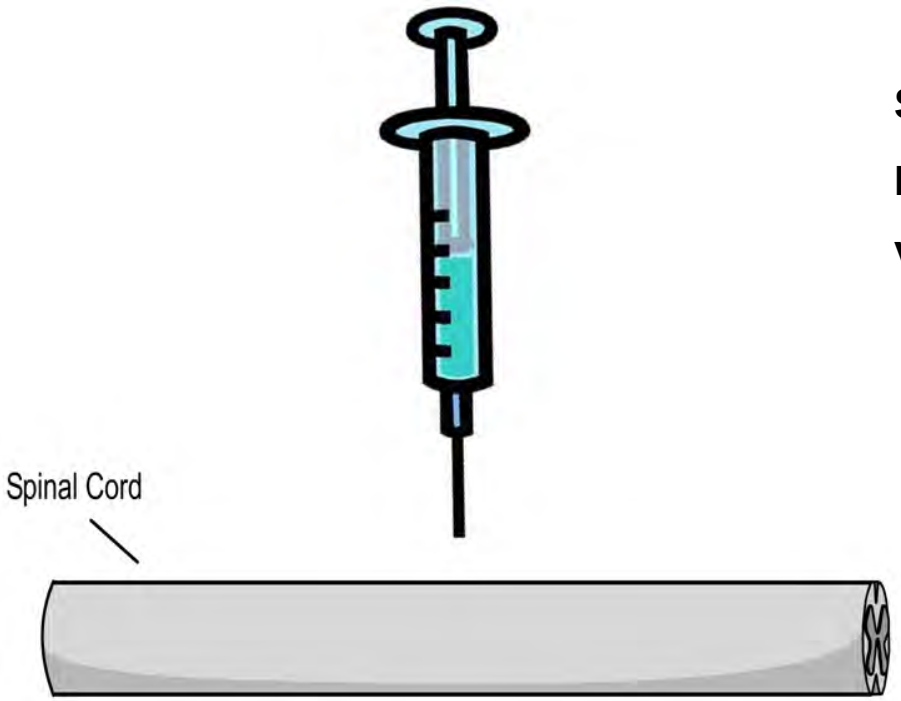
- Stimulation of cell proliferation
- Prevention of apoptosis
- Cell migration
- Inflammation
- Platelet aggregation, etc..

Atherosclerosis, cancer and pulmonary fibrosis.

In vitro studies: neurite retraction, neuronal death and microglial activation.

Lysophosphatidic Acid

LPA INTRASPINAL INJECTION

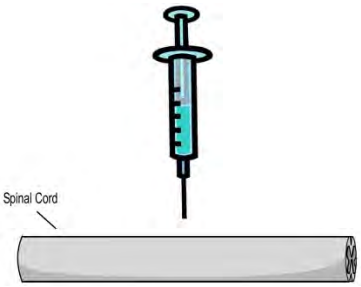


Strain: C57Bl/6

Level: T11

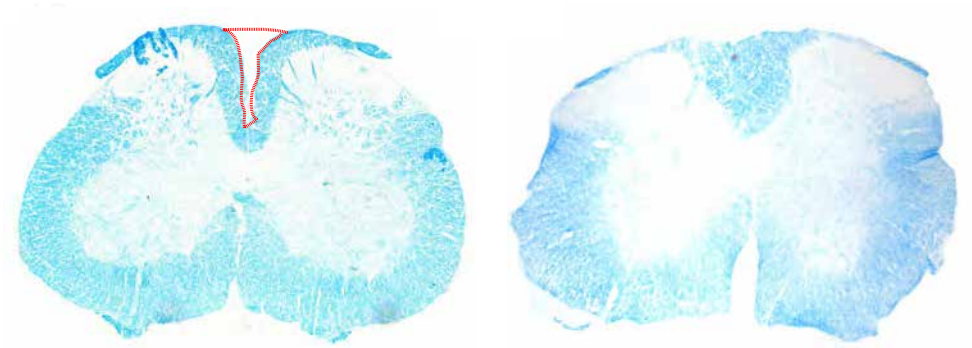
Volume: 1µl containing 5nmoles of LPA

Lysophosphatidic Acid



Epicenter

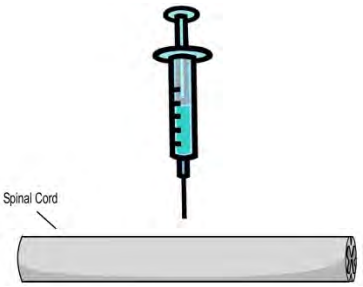
1800 μm Rostral

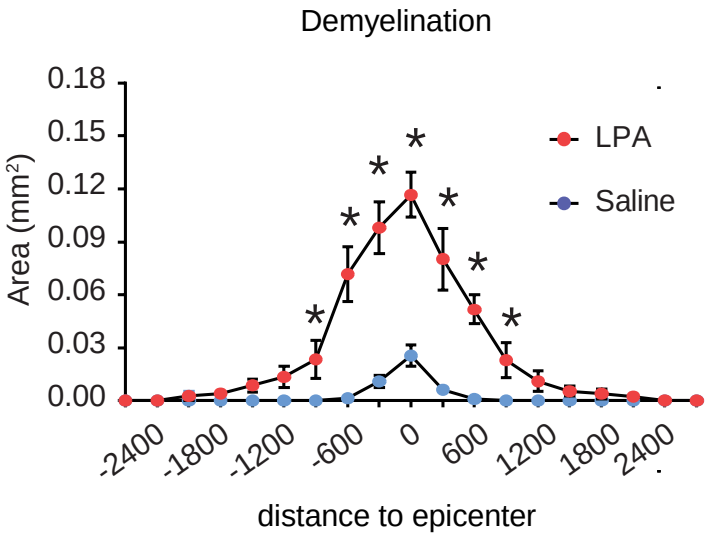


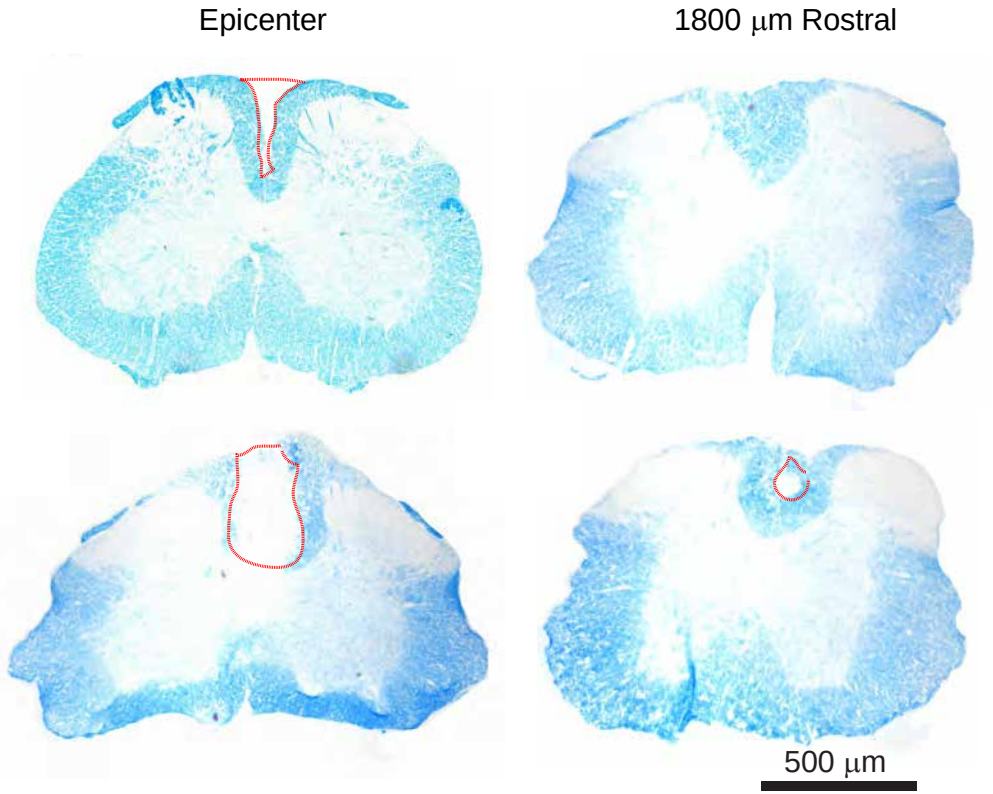
500 μm

Santos-Nogueira et al., 2015 J Neurosci

Lysophosphatidic Acid

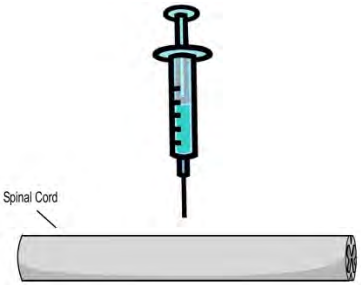


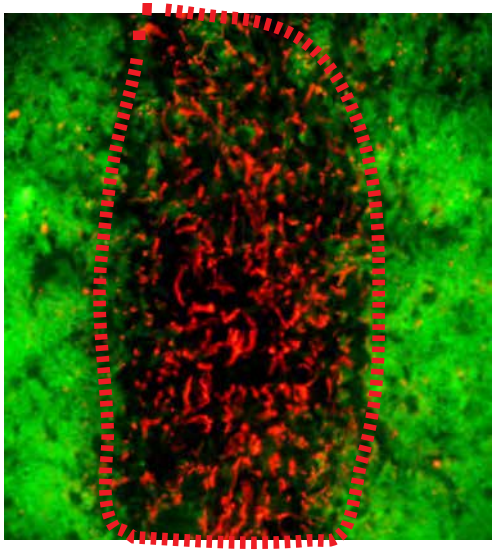




Santos-Nogueira et al., 2015 J Neurosci

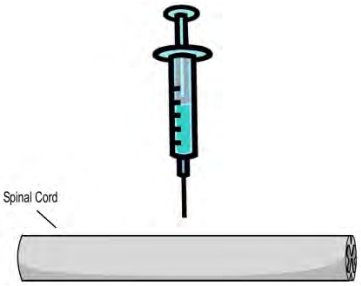
Lysophosphatidic Acid

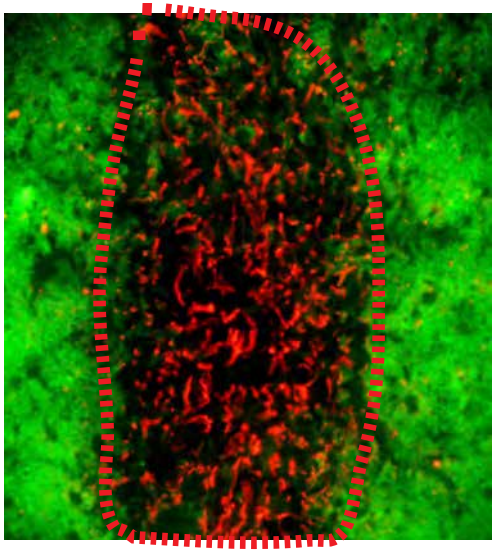


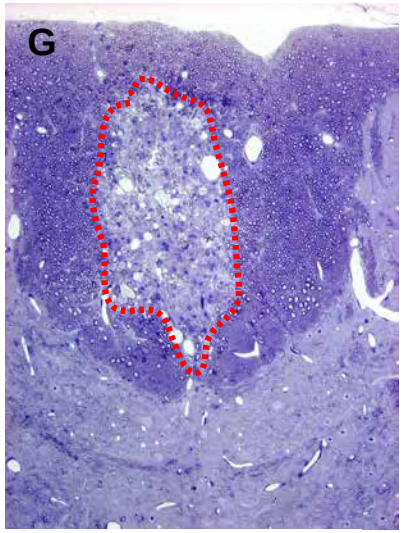


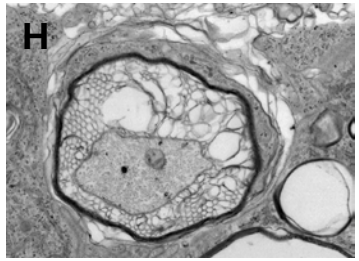
Santos-Nogueira et al., 2015 J Neurosci

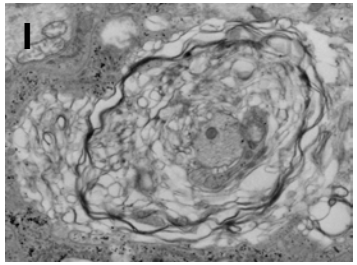
Lysophosphatidic Acid

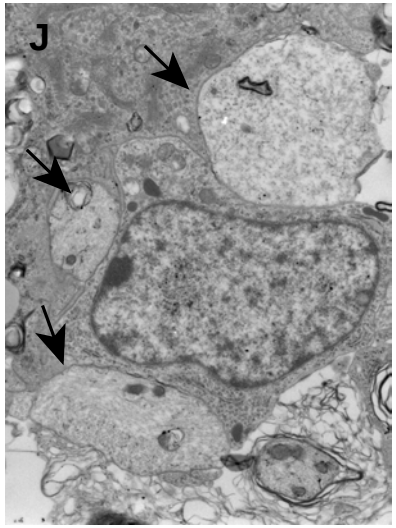






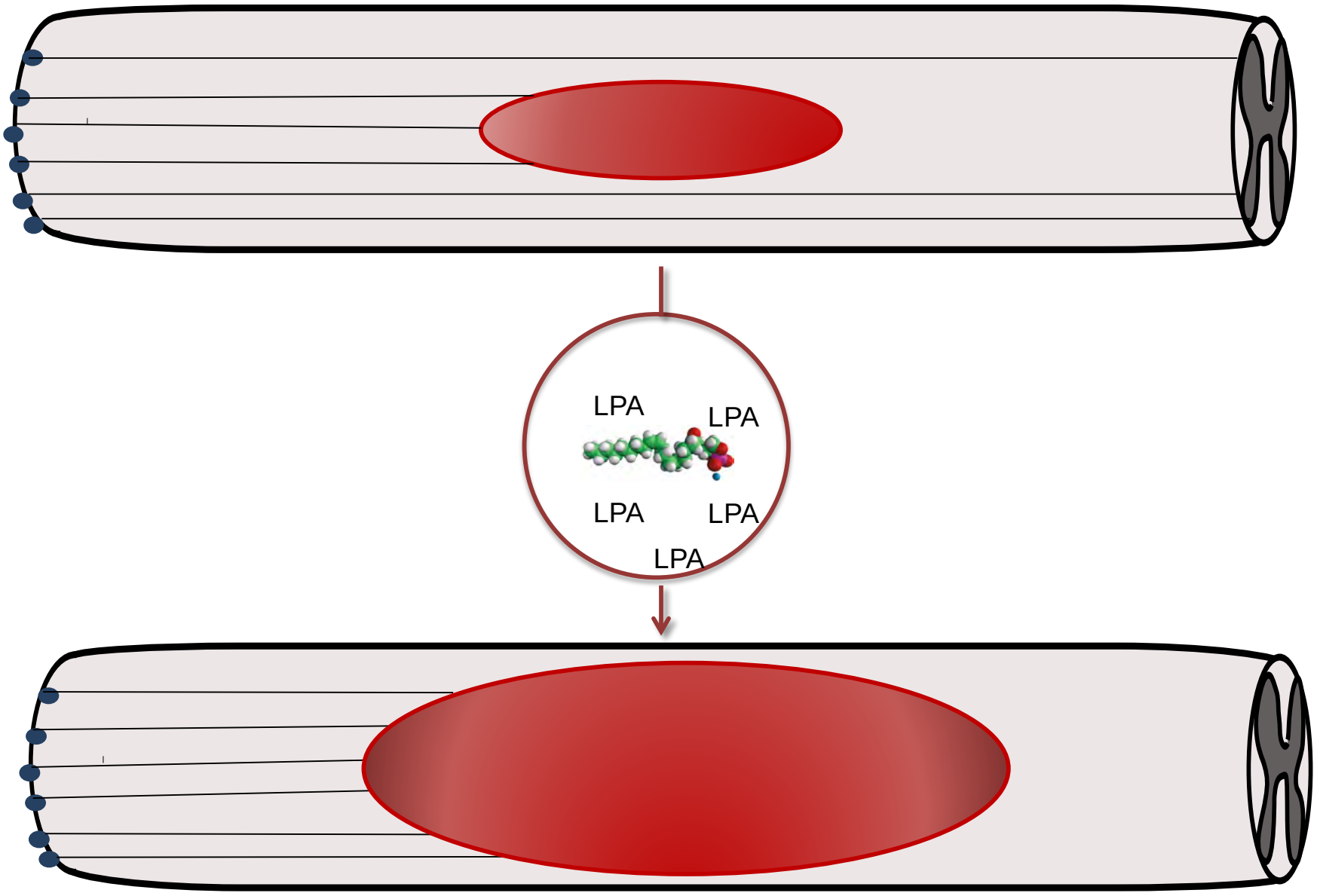




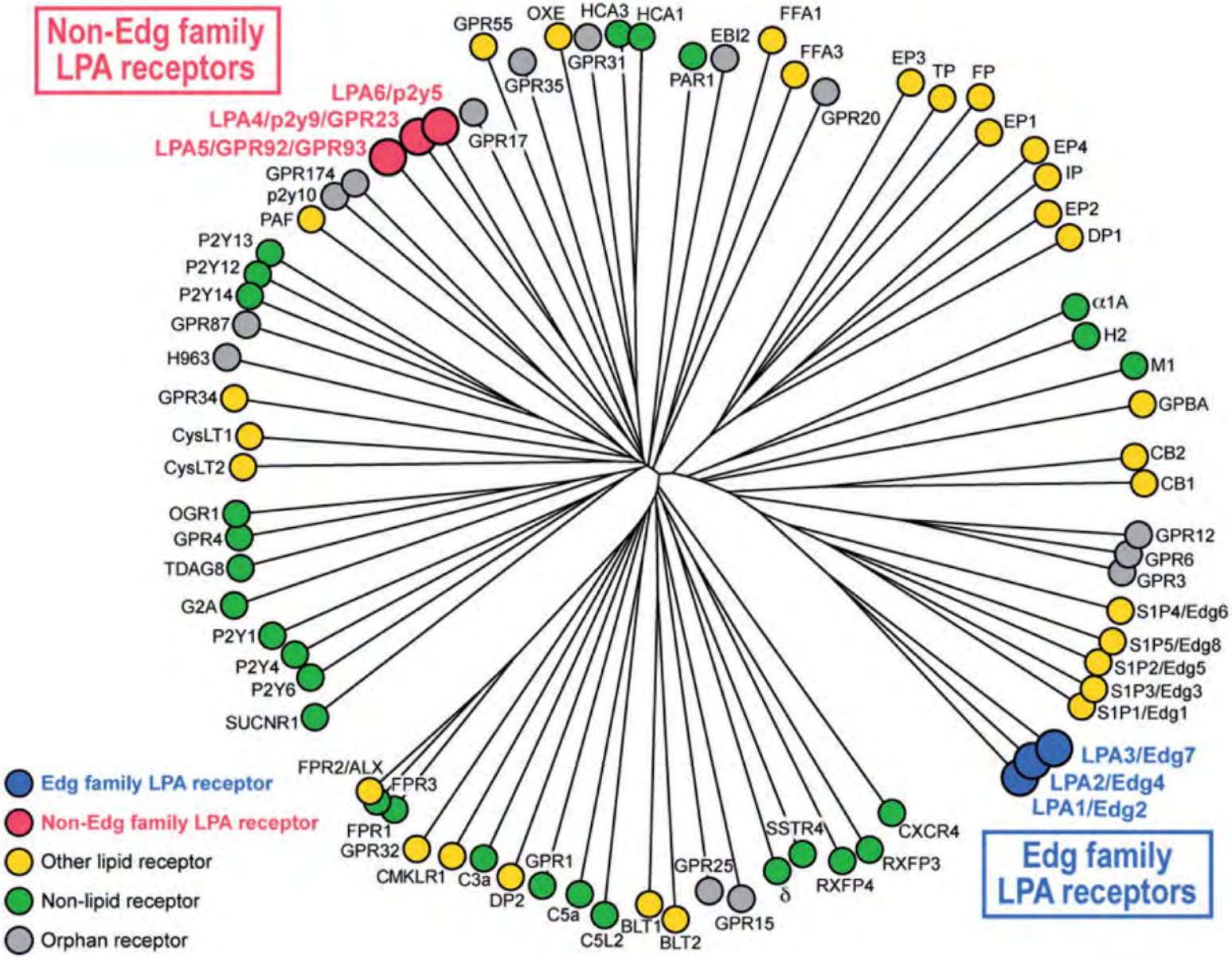


Santos-Nogueira et al., 2015 J Neurosci

Lysophosphatydic Acid

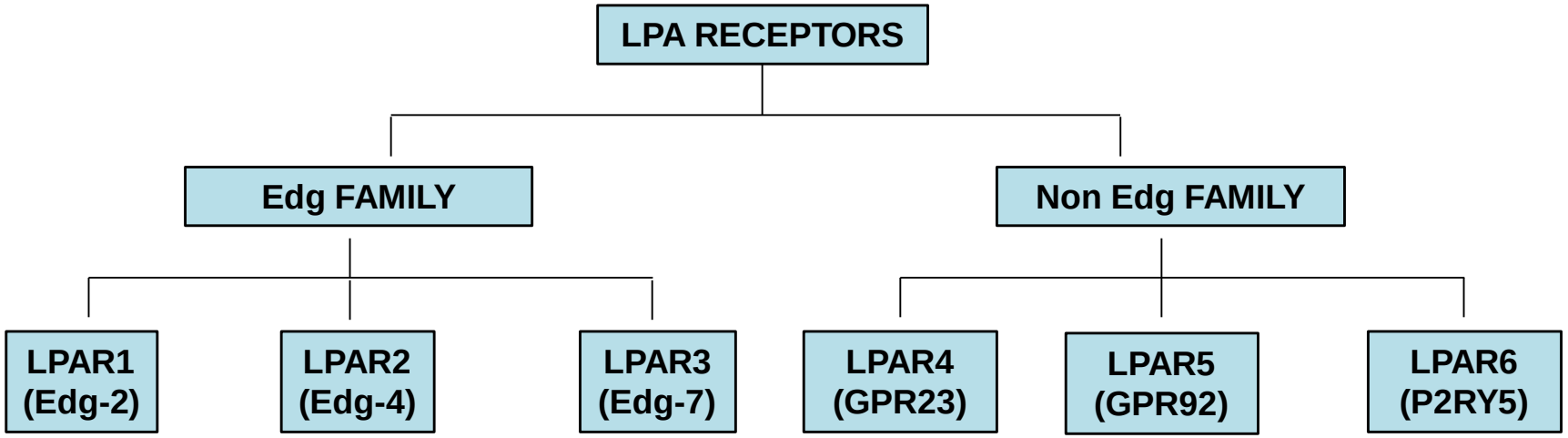


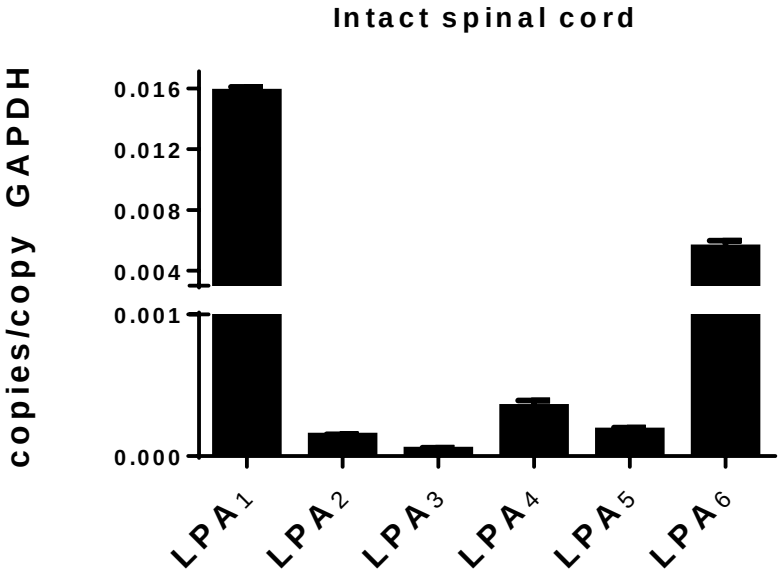
Lysophosphatidic Acid



Yamagida and Ishii, 2011, J Biochem

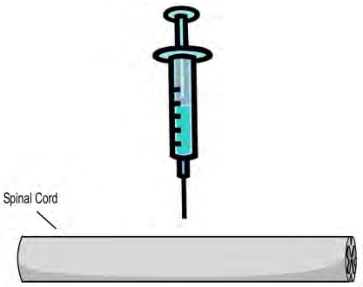
Lysophosphatydic Acid



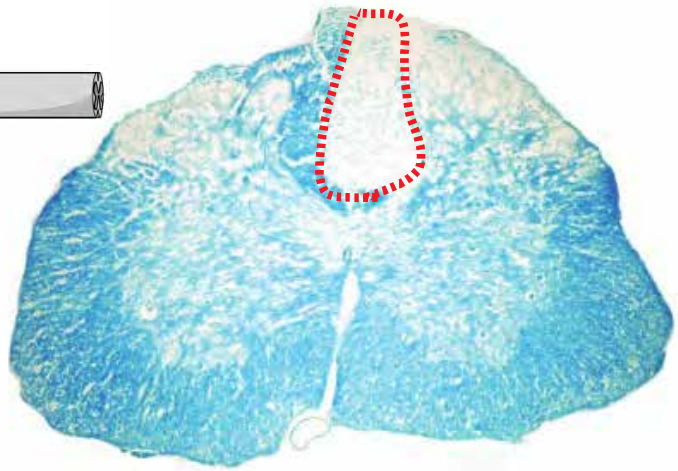


Santos-Nogueira et al., 2015 J Neurosci

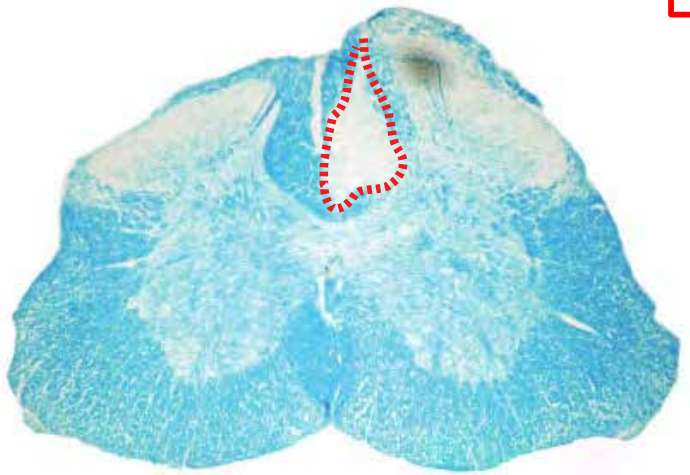
LPA receptor 1

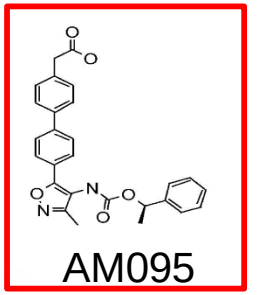


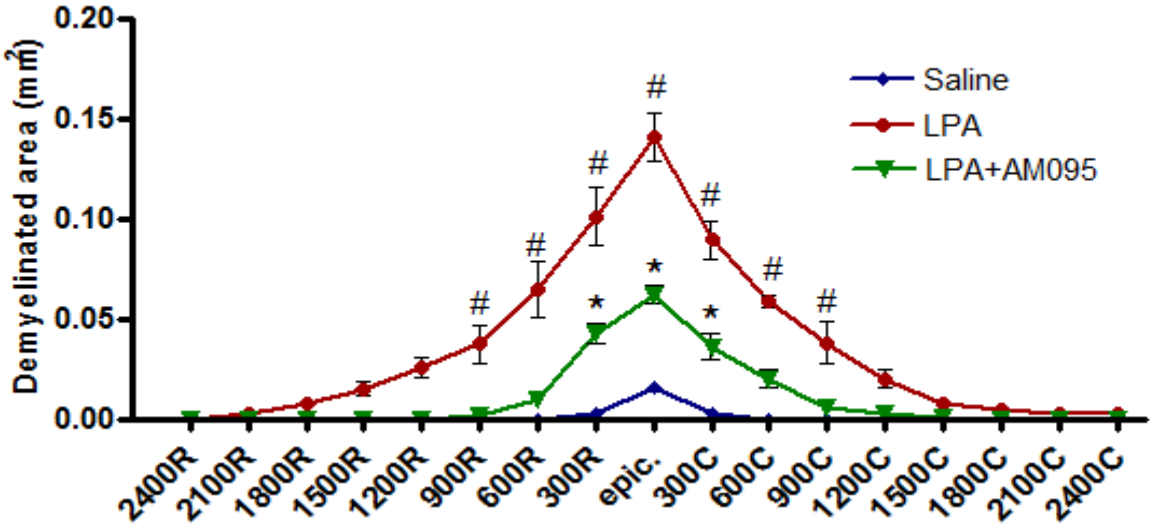
LPA



LPA + AM095



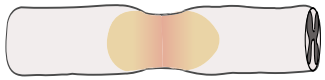


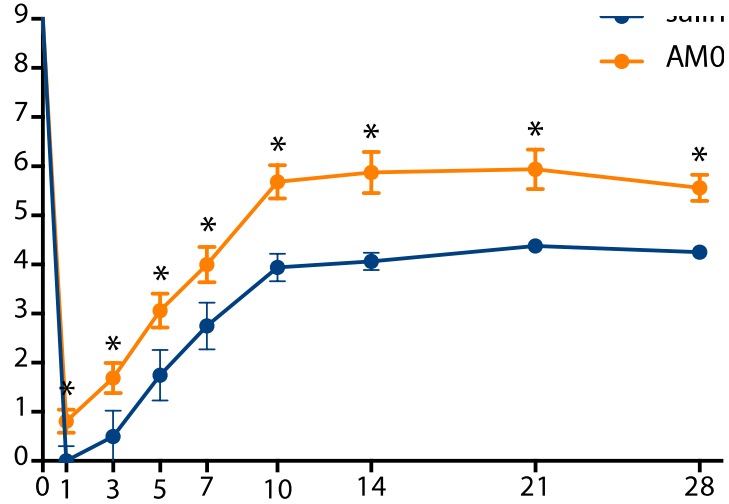


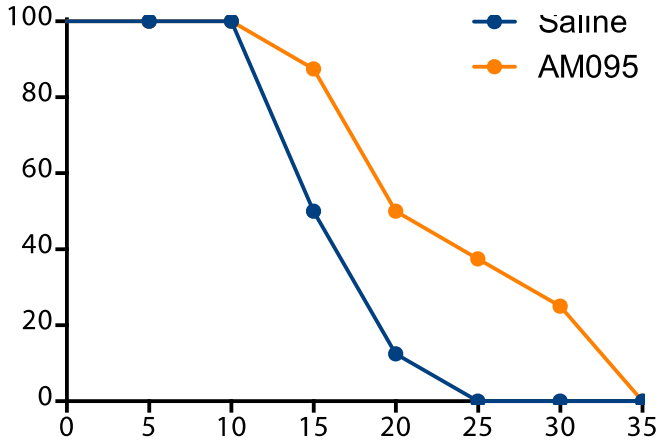
# p < 0.05 LPA *vs.* saline
* P < 0.05 AM095 *vs.* LPA

Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 1



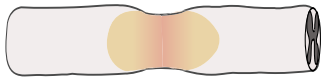


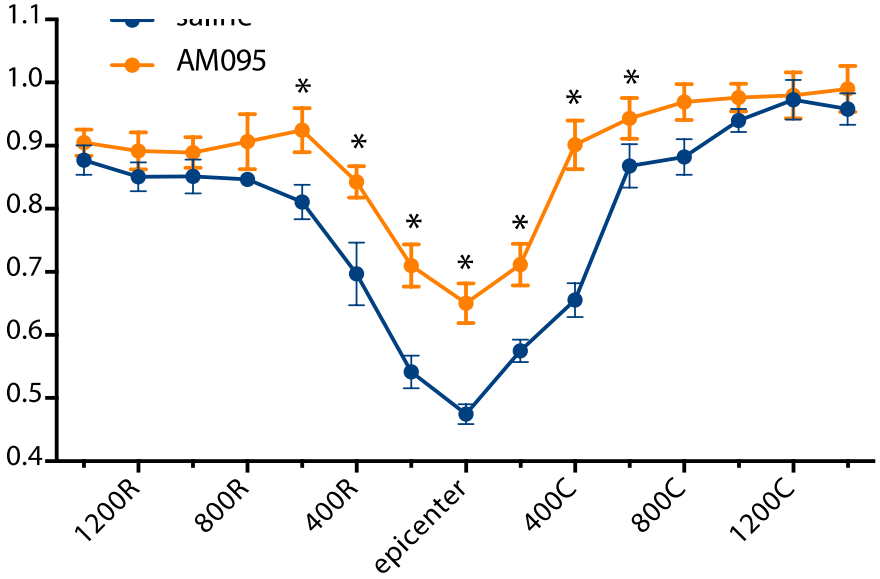


* $p < 0.05$

Santos-Nogueira et al., 2015 J Neurosci

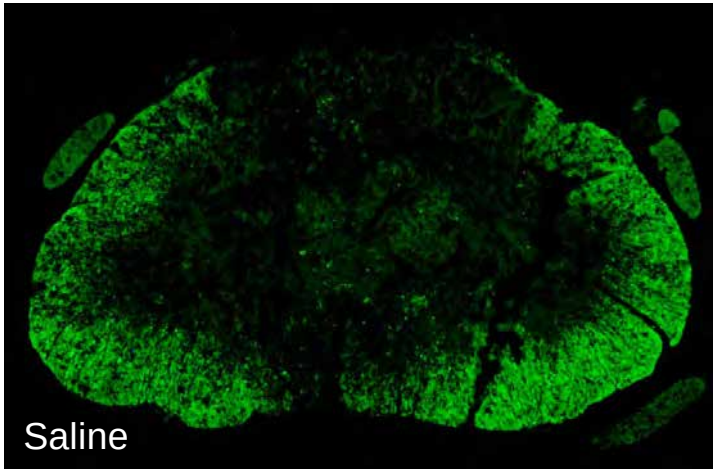
LPA receptor 1

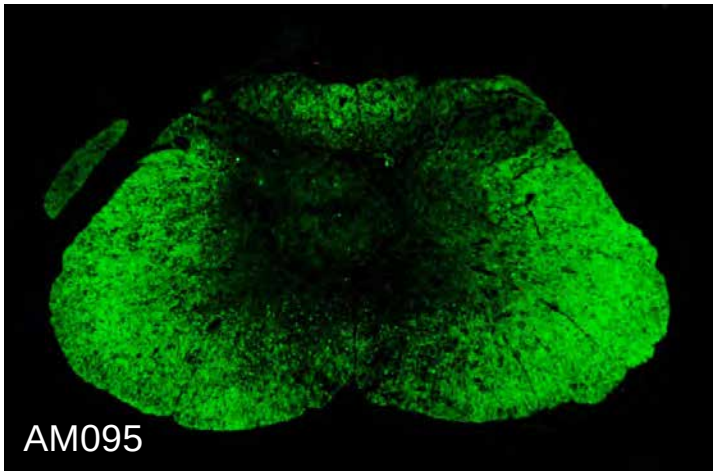




* $p < 0.05$

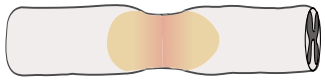
Epicenter

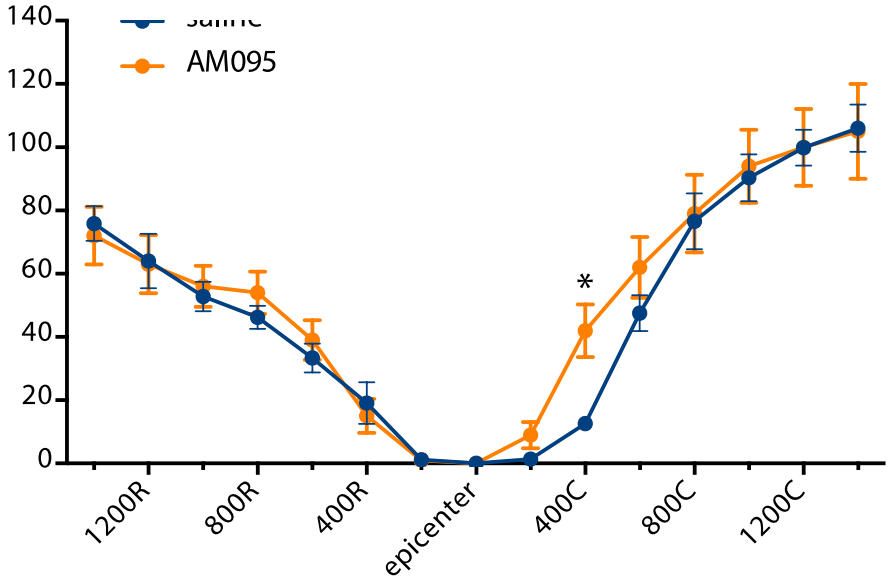




Santos-Nogueira et al., 2015 J Neurosci

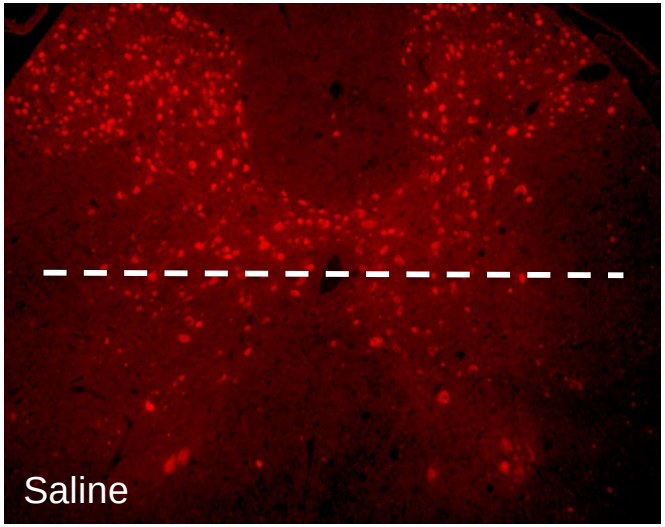
LPA receptor 1

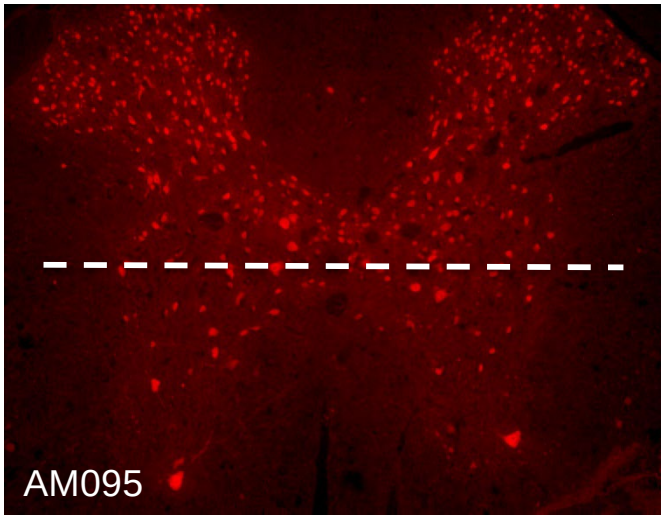




* $p < 0.05$

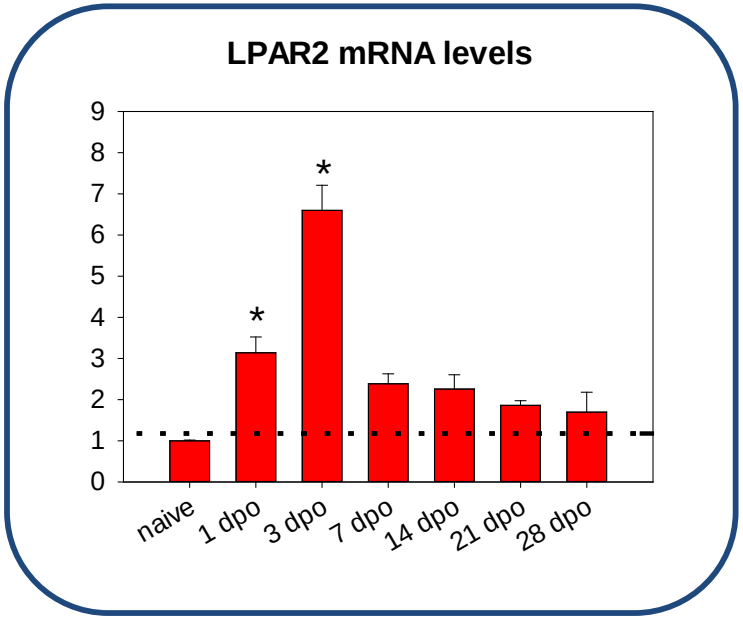
600 µm Rostral

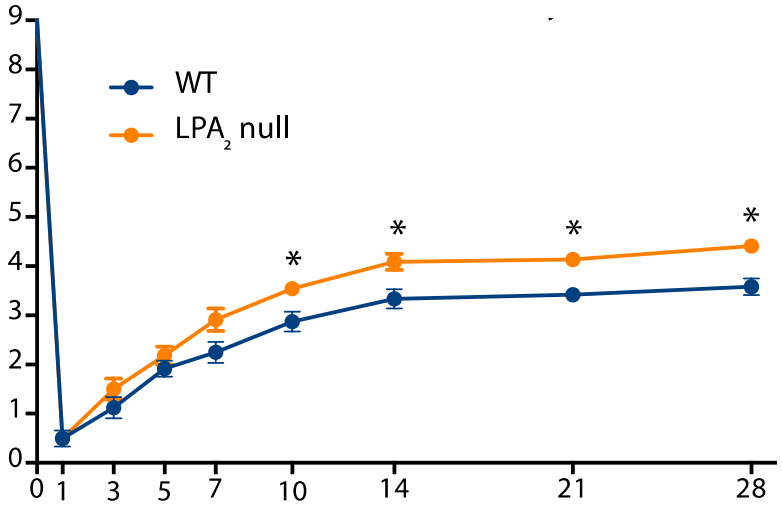


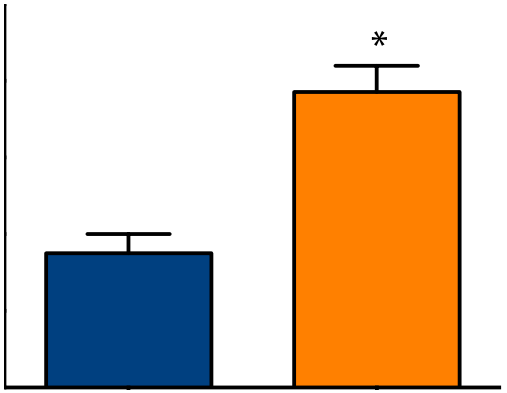


Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 2

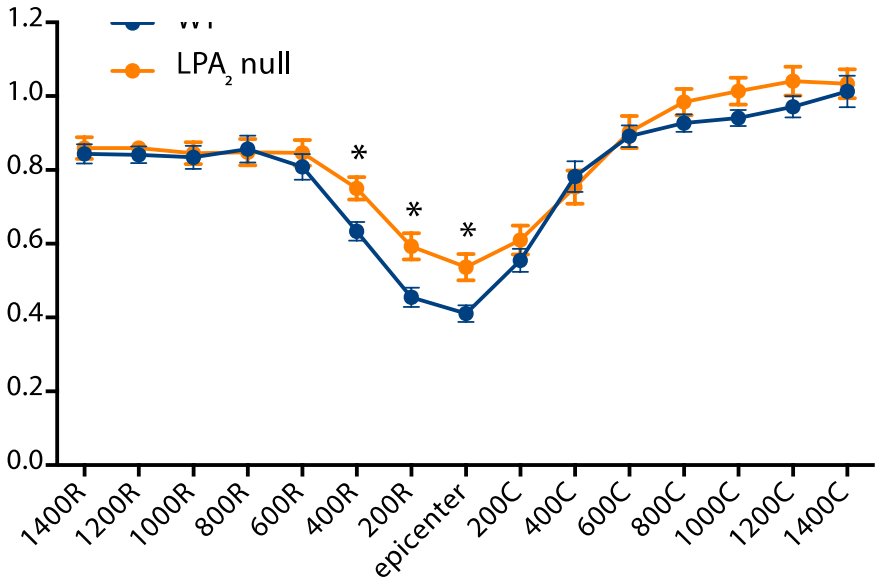


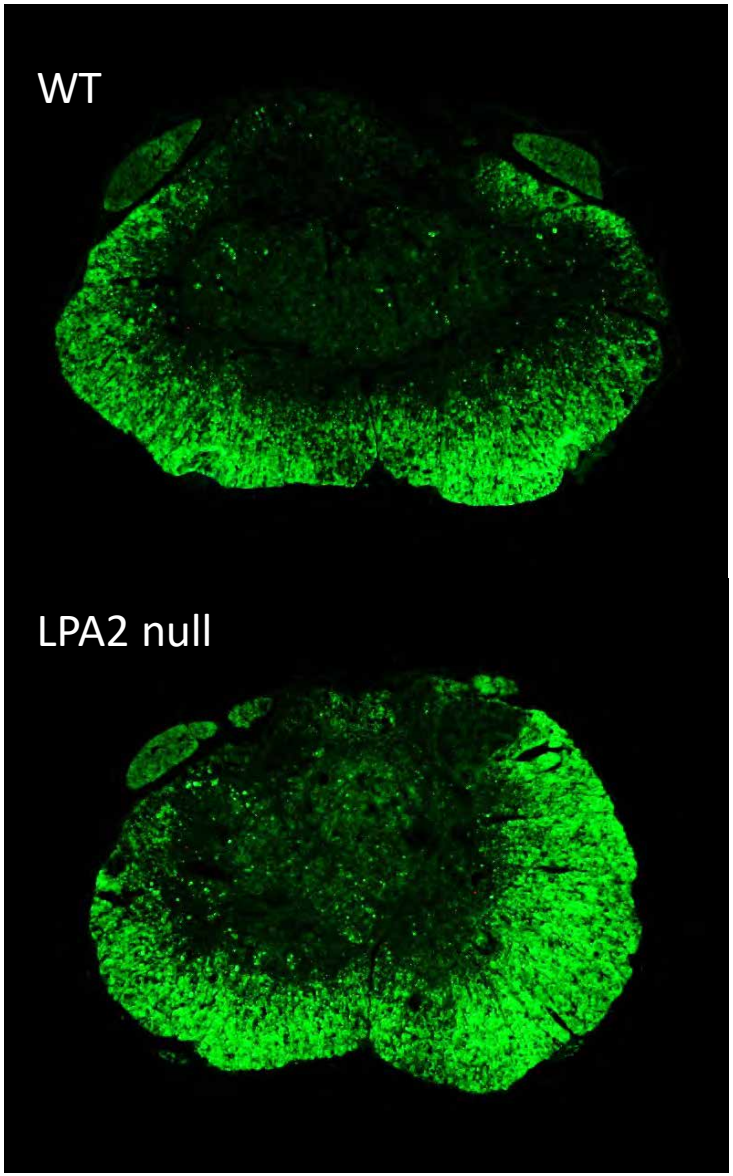




Lopez-Serrano et al., Brain Beh Immun 2019

LPA receptor 2



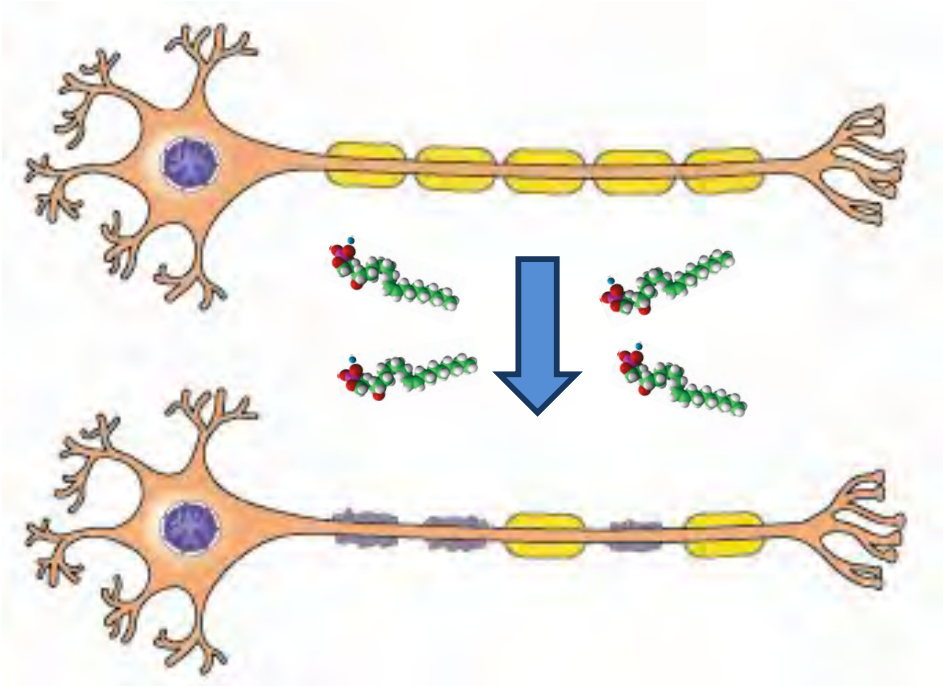


* $p < 0.05$

Lopez-Serrano et al., Brain Beh Immun 2019

LPA receptor 1

How does LPA-LPA_1 signaling induce demyelination in the spinal cord?



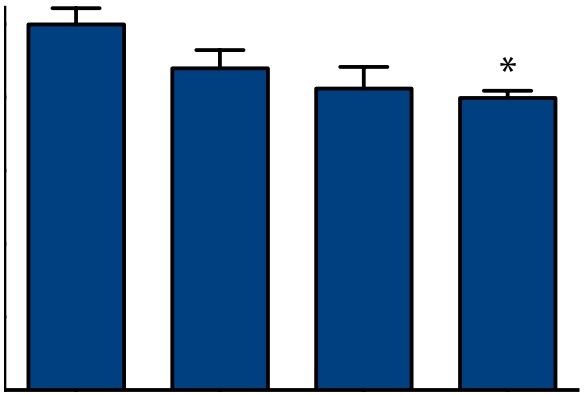
Lysophosphatidic acid

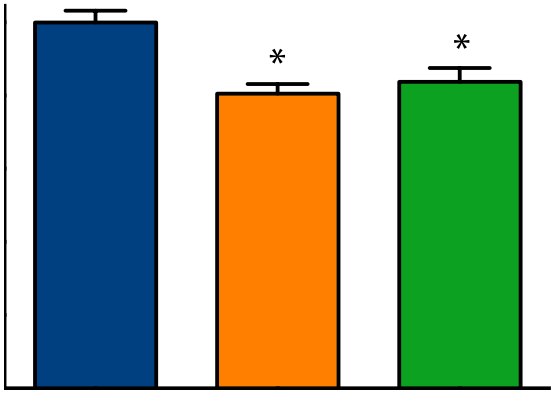
OLIGODENDROCYTE SURVIVAL: *in vitro* studies

OL ENRICHED CULTURE

LPA exposure (for 24h)

Control
LPA (0.01, 0.1, 1μM)



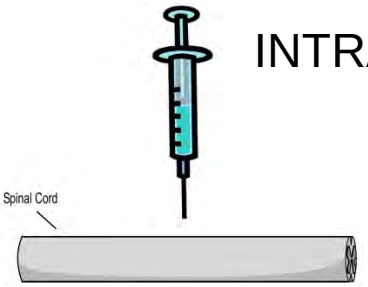


* $p < 0.05$

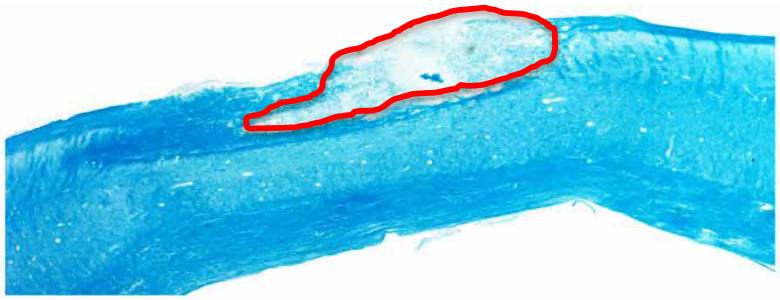
Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 1

INTRASPINAL INJECTION OF LYSOPHOSPHATIDIC ACID

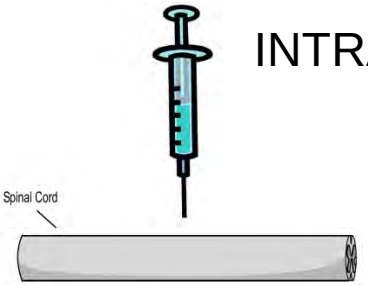


Myelin
(LFB)



Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 1



INTRASPINAL INJECTION OF LYSOPHOSPHATIDIC ACID

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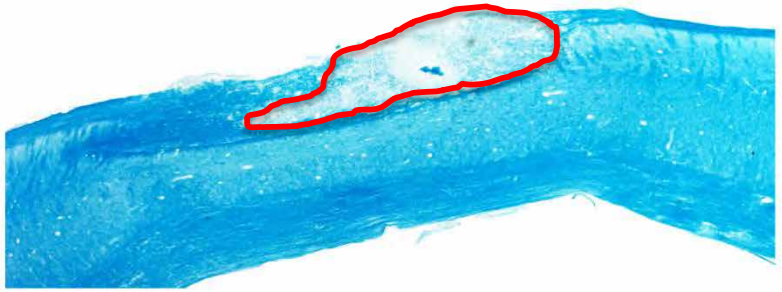
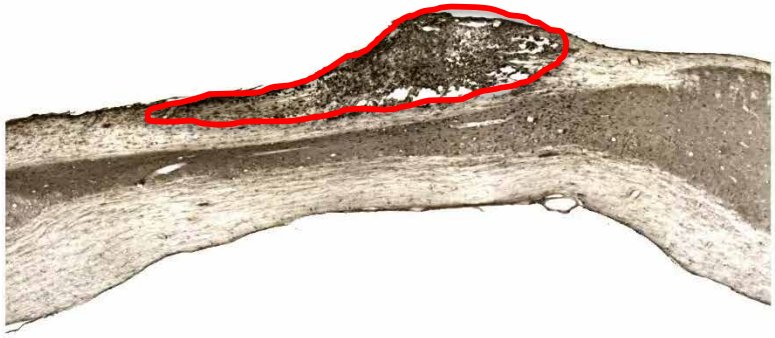
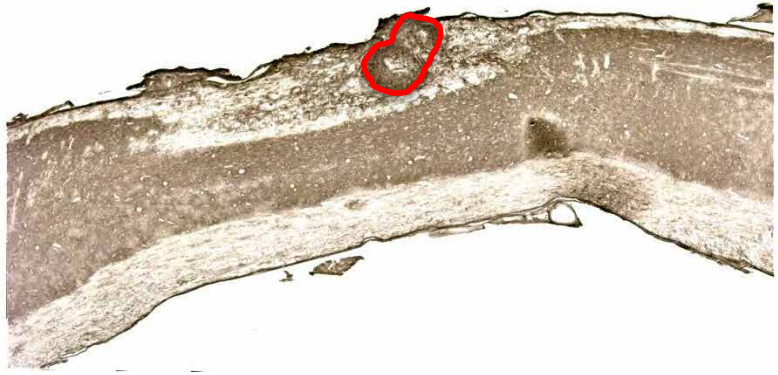


Microglia and
macrophages
(Iba1)

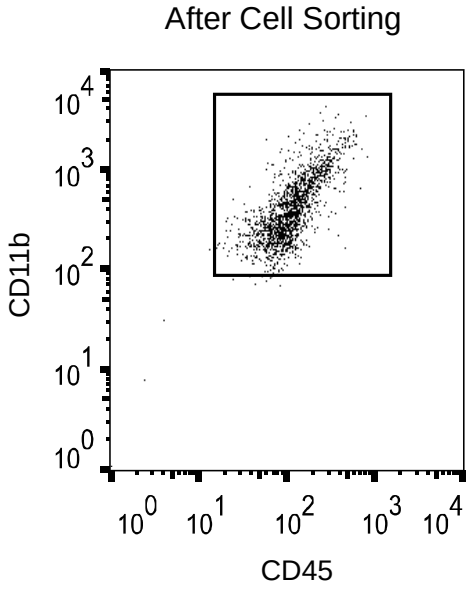


Astrocytes
(GFAP)

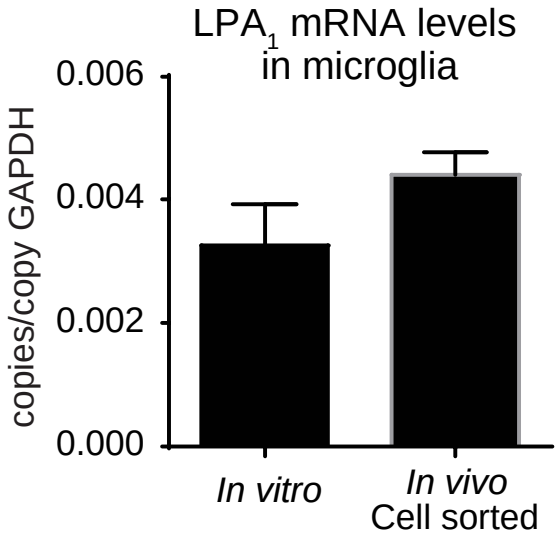


Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 1



CD11b immunopanning ▸ Cell sorting

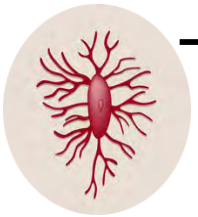


Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 1

OLIGODENDROCYTE SURVIVAL: *in vitro* studies

MICROGLIA CULTURE



→ Generation of conditioned medium

- Control
- 1μM LPA
- 1μM LPA + 12.5μM AM095

→ OL exposure to conditioned medium (for 24h)

Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 1

OLIGODENDROCYTE SURVIVAL: *in vitro* studies

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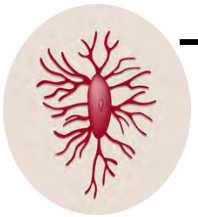

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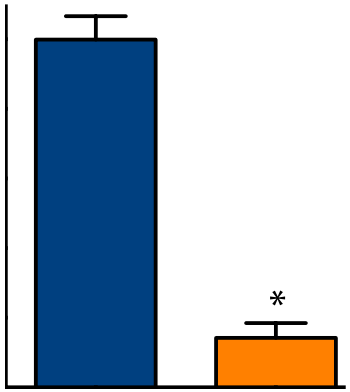
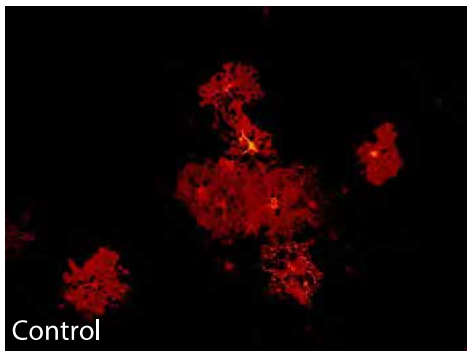
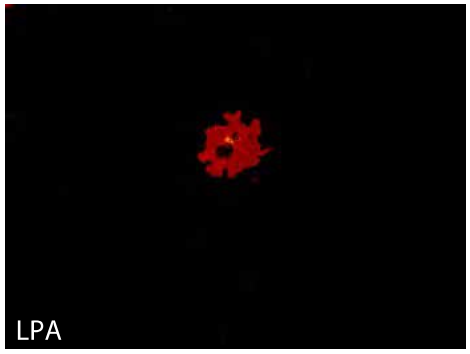






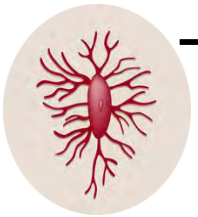
* $p < 0.05$ vs. control
\# $p < 0.05$ vs. LPA

Santos-Nogueira et al., 2015 J Neurosci

LPA receptor 1

OLIGODENDROCYTE SURVIVAL: *in vitro* studies

MICROGLIA CULTURE



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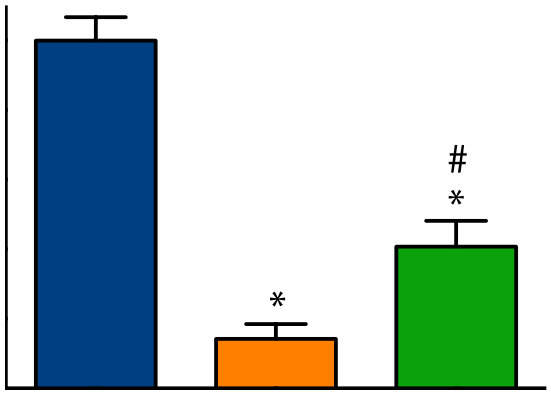

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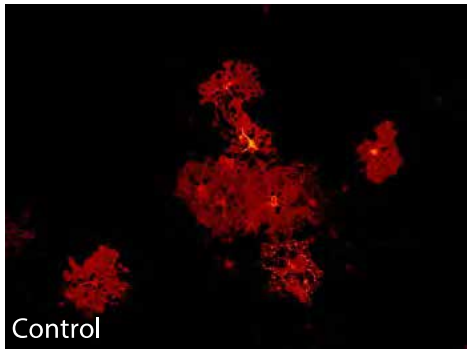
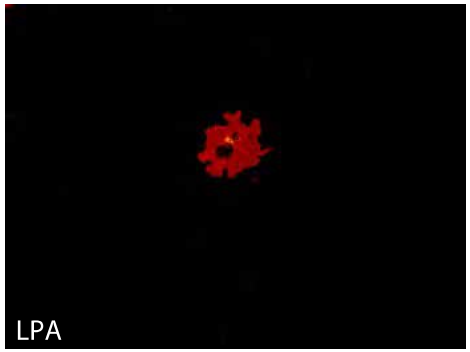
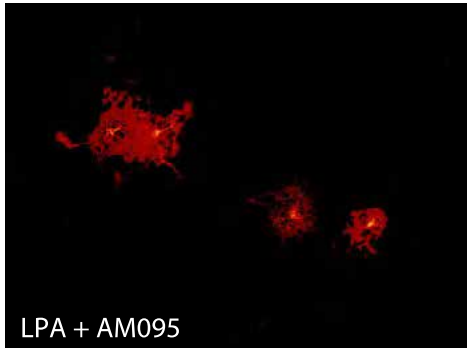
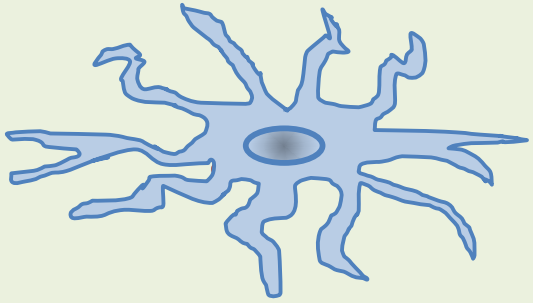
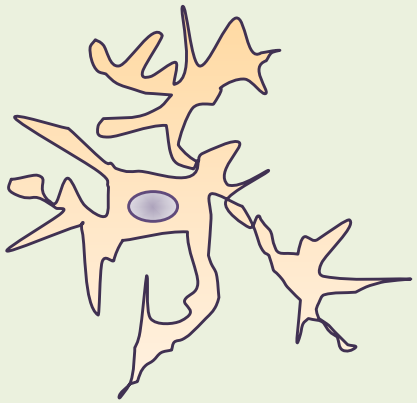
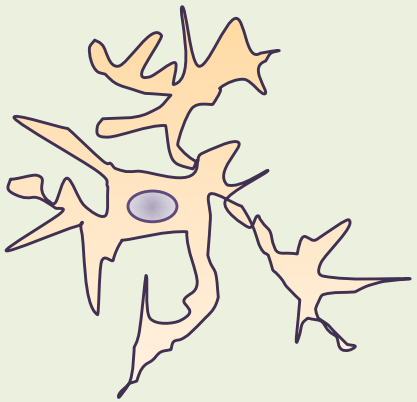
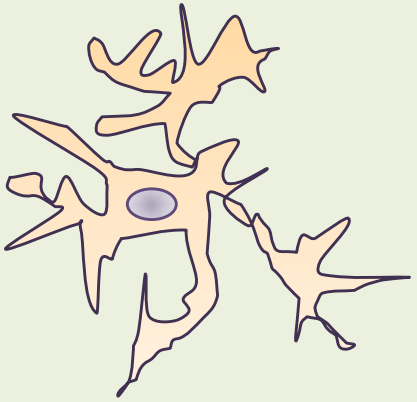


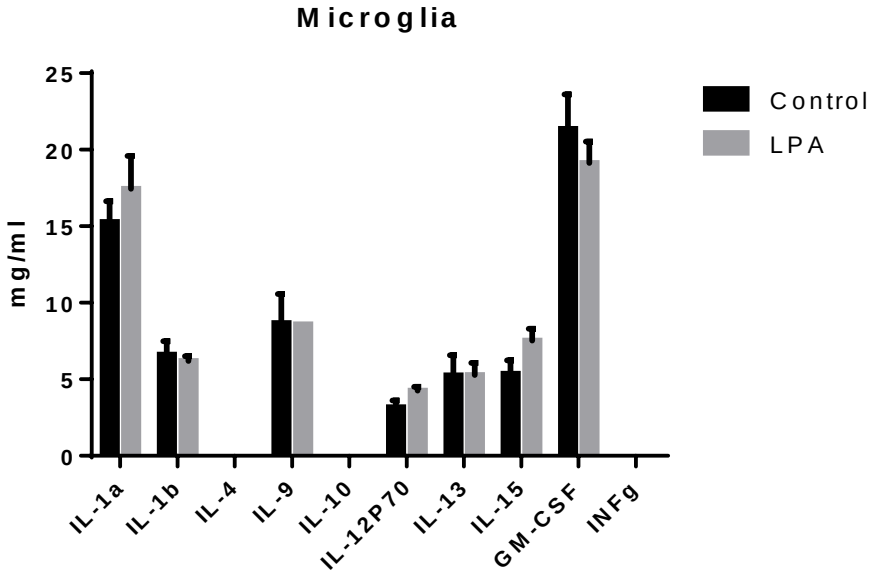


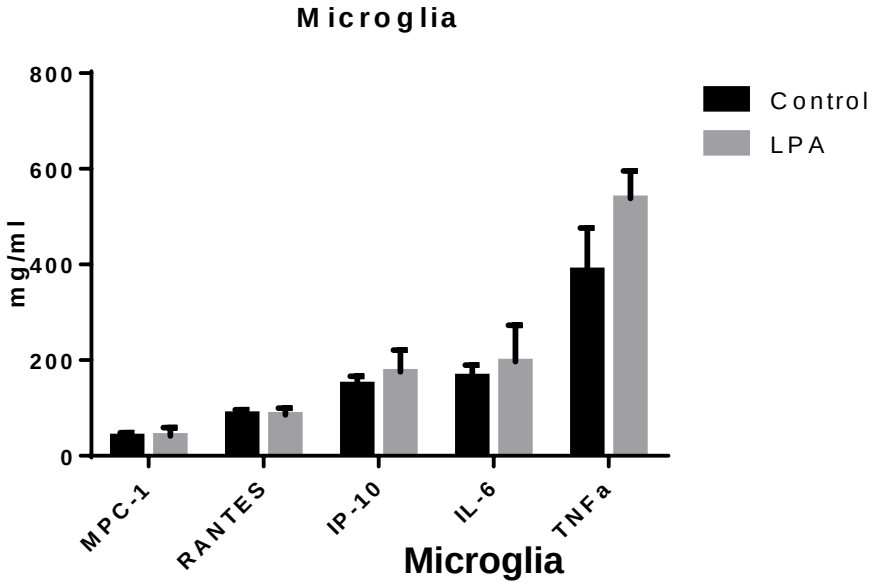


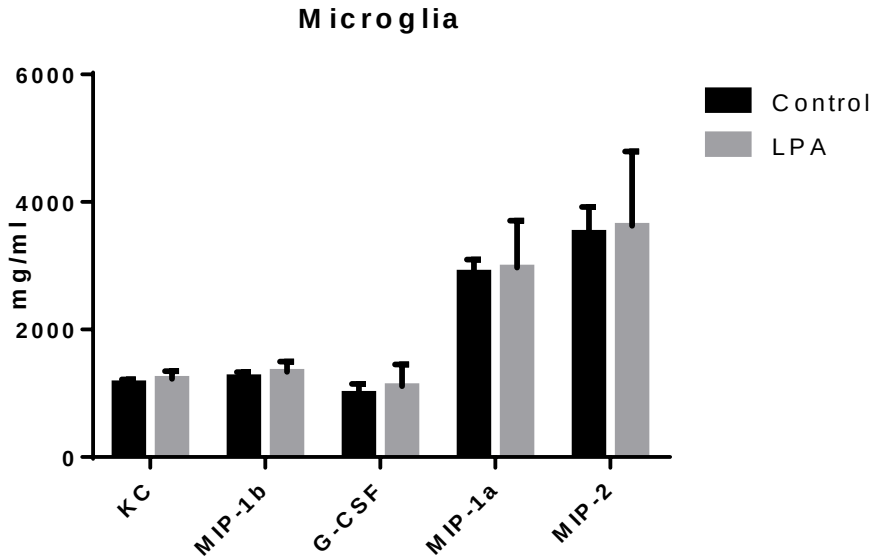
Santos-Nogueira et al., 2015 J Neurosci

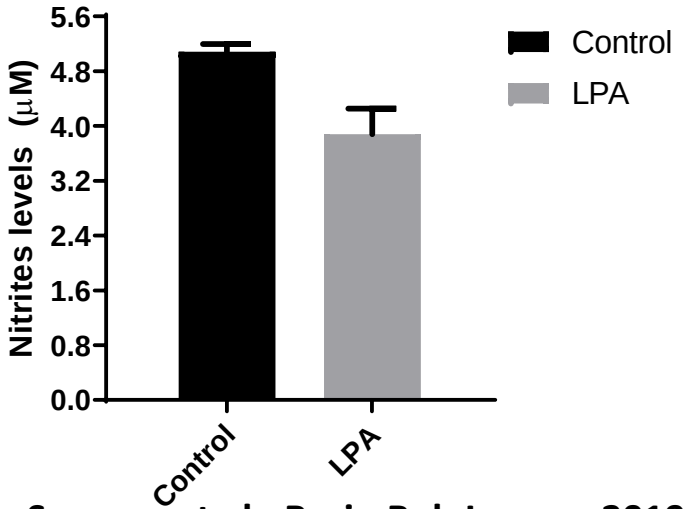





How does microglia induce oligodendrocyte cell death upon LPA stimulation?









Lopez-Serrano et al., Brain Beh Immun 2019

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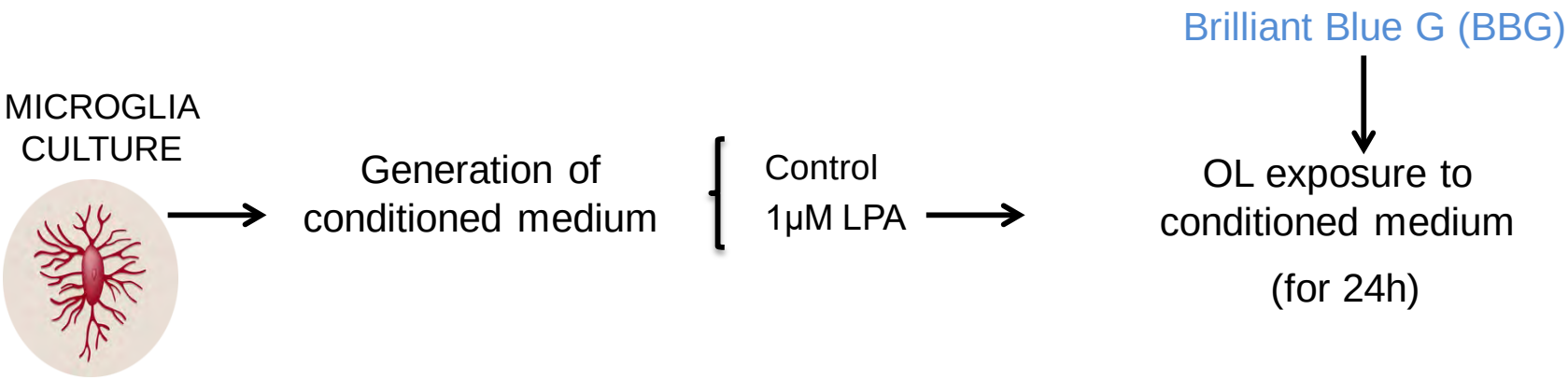
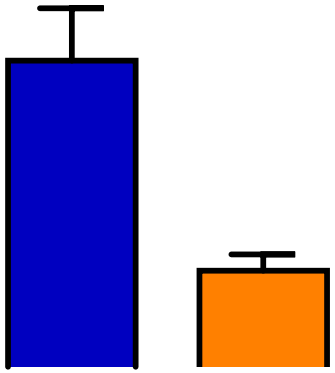




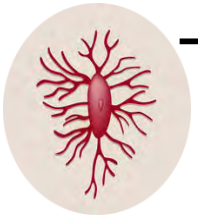
* $p < 0.001$ vs. control
\# $p < 0.05$ vs. LPA vehicle

Lopez-Serrano et al., Brain Beh Immun 2019

How does microglia induce oligodendrocyte cell death upon LPA stimulation?

Brilliant Blue G (BBG)

MICROGLIA CULTURE

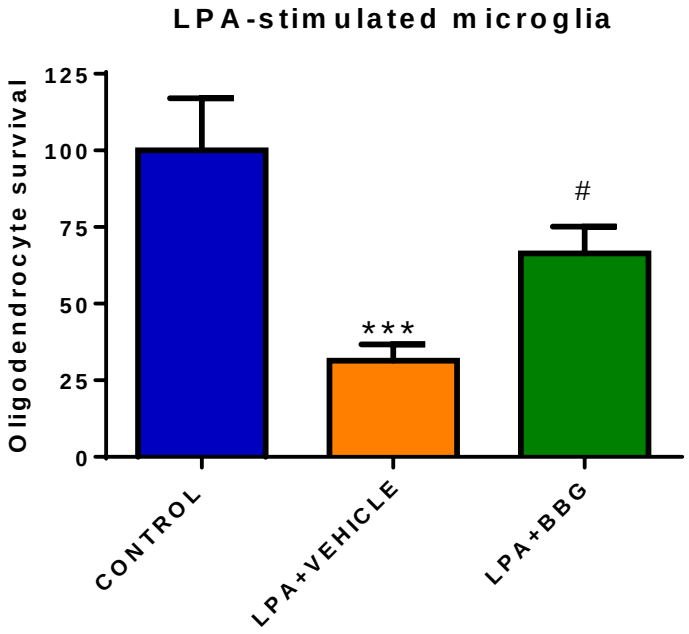


Generation of conditioned medium

Control
1μM LPA

OL exposure to conditioned medium

(for 24h)



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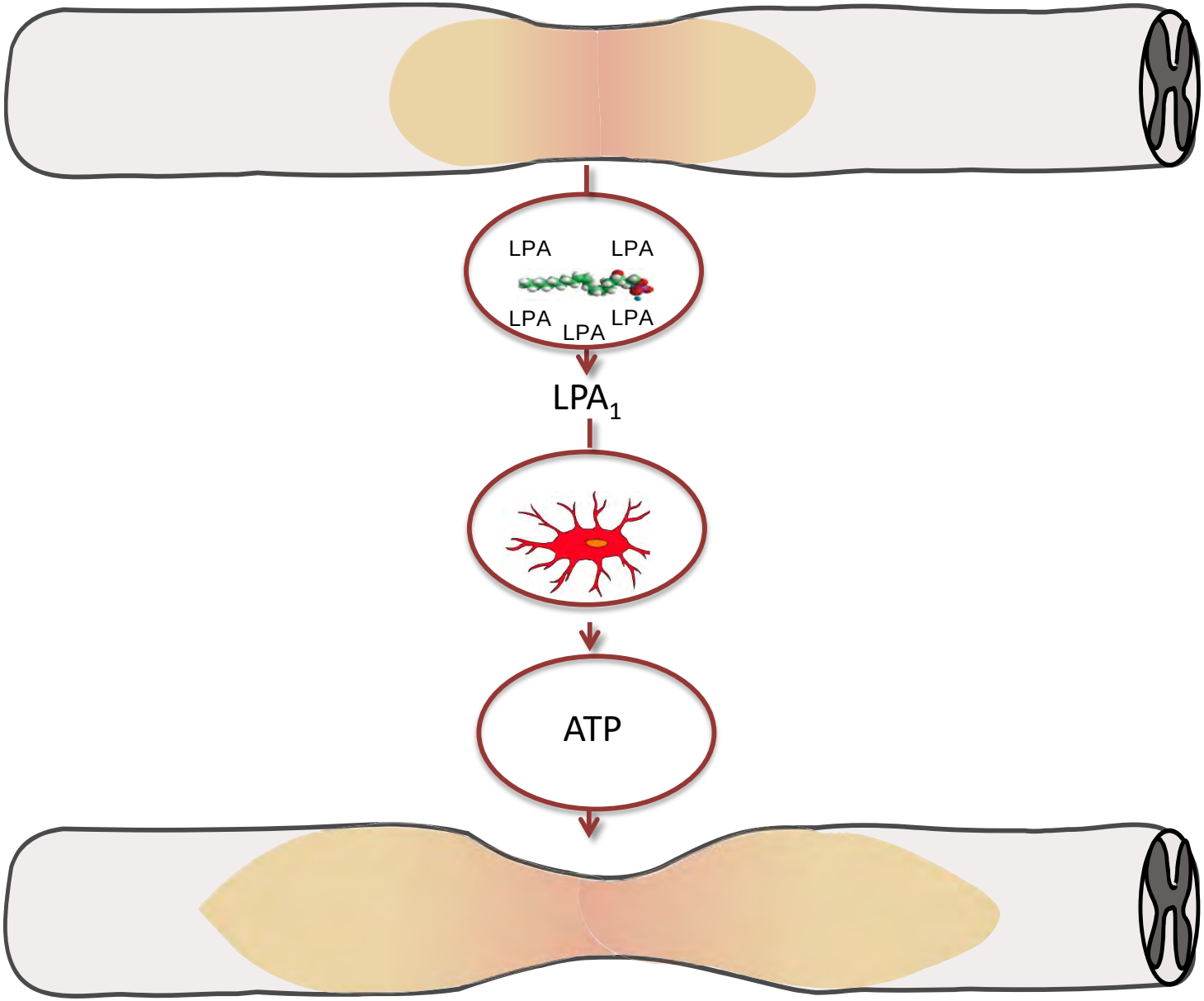
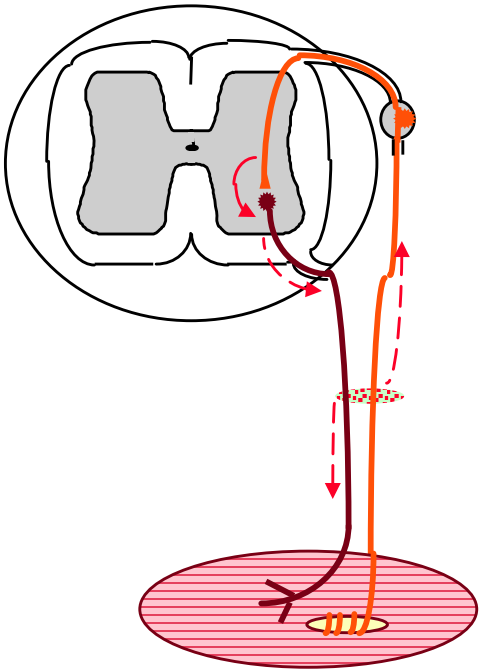
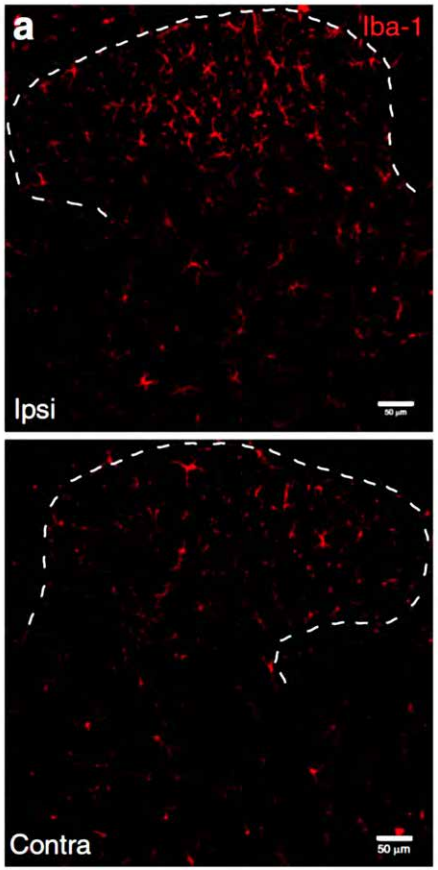
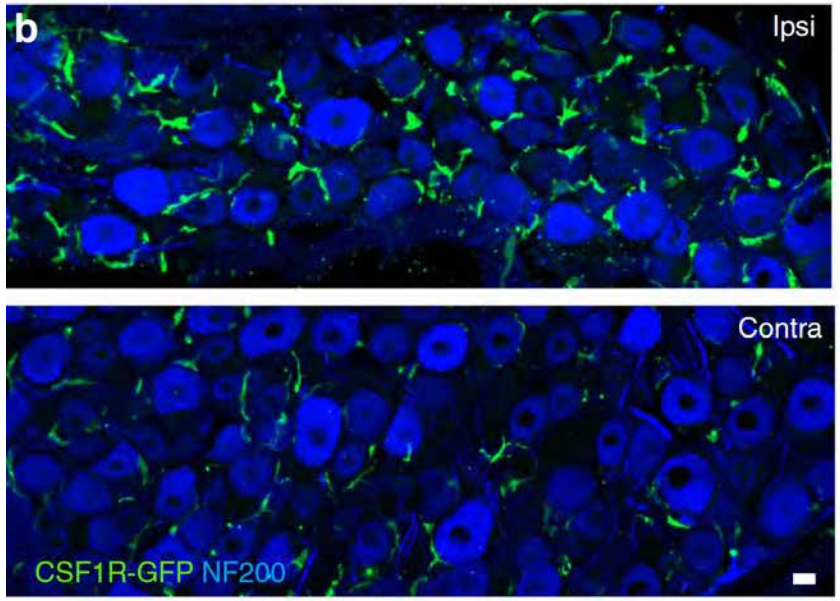
Lopez-Serrano et al., Brain Beh Immun 2019

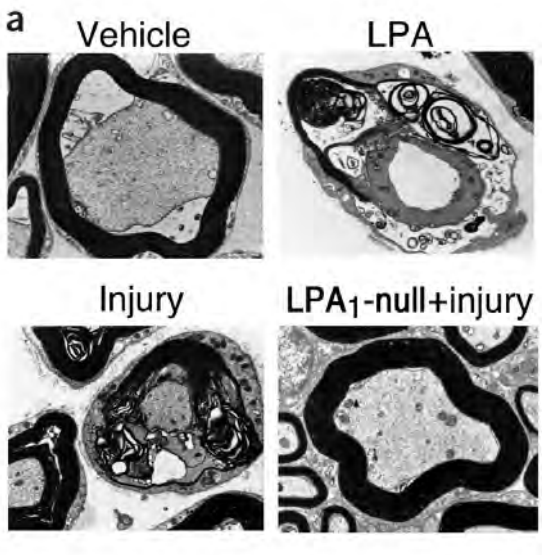

LPA
LPA
LPA
LPA
LPA
LPA1
ATP

LPA receptor 1 after sciatic nerve injury



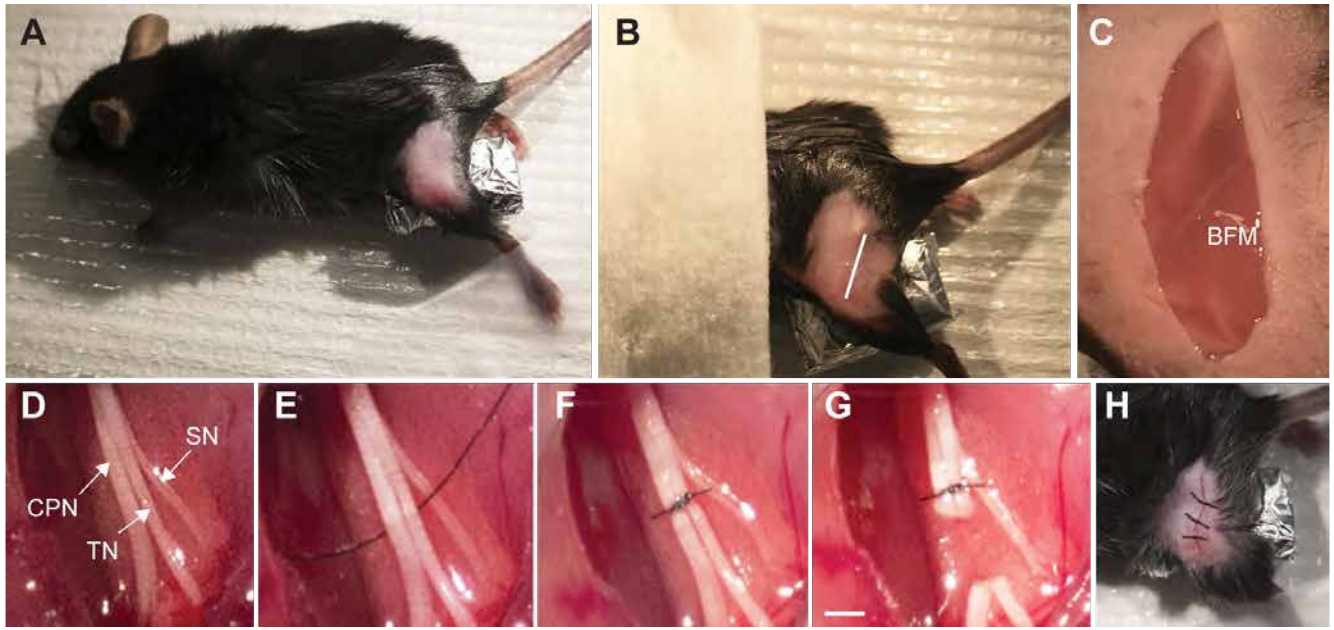


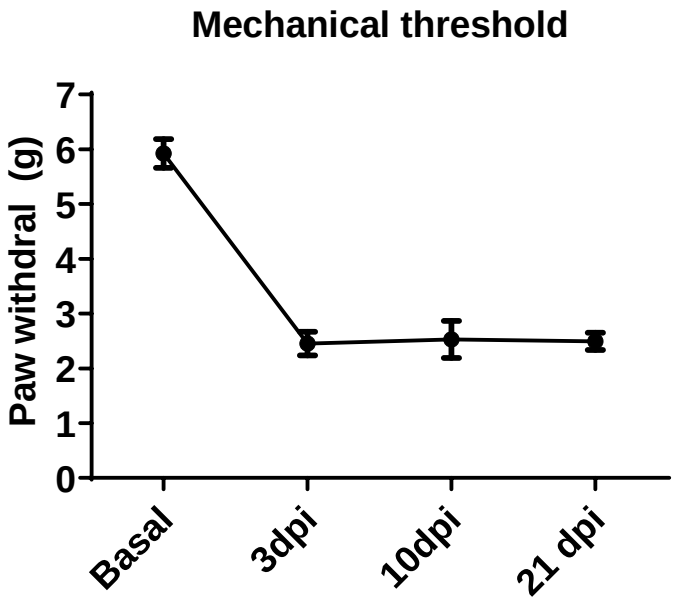


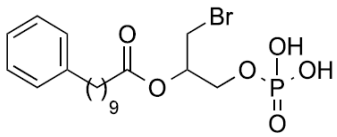


Yu et al., 2020; Unue et al., 2007

LPA receptor 1 after sciatic nerve injury

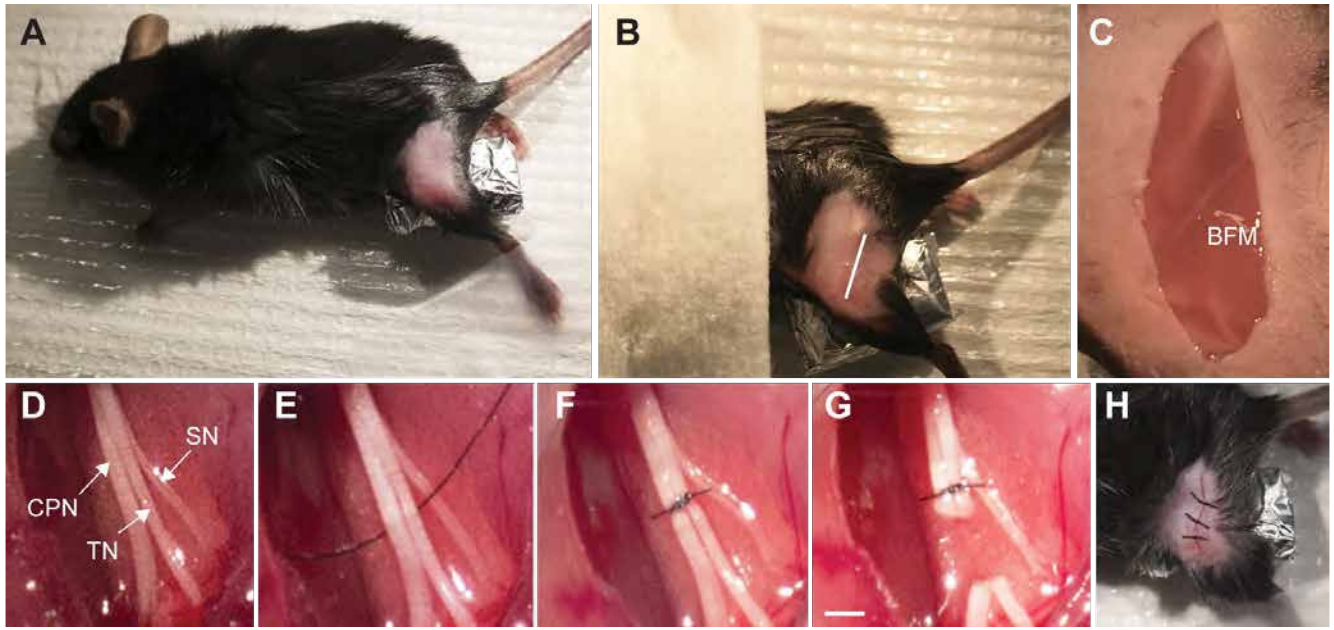


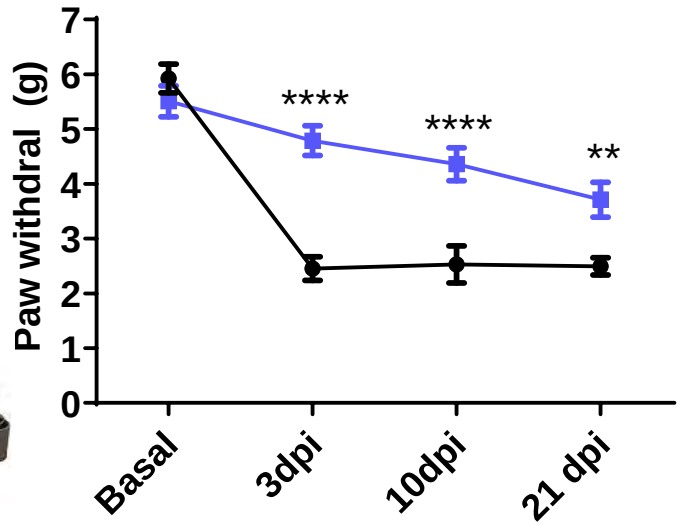




González-Gil et al., 2021

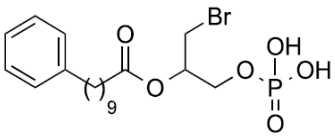
LPA receptor 1 after sciatic nerve injury





(S)-17 LPA1 antagonist

Vehicle



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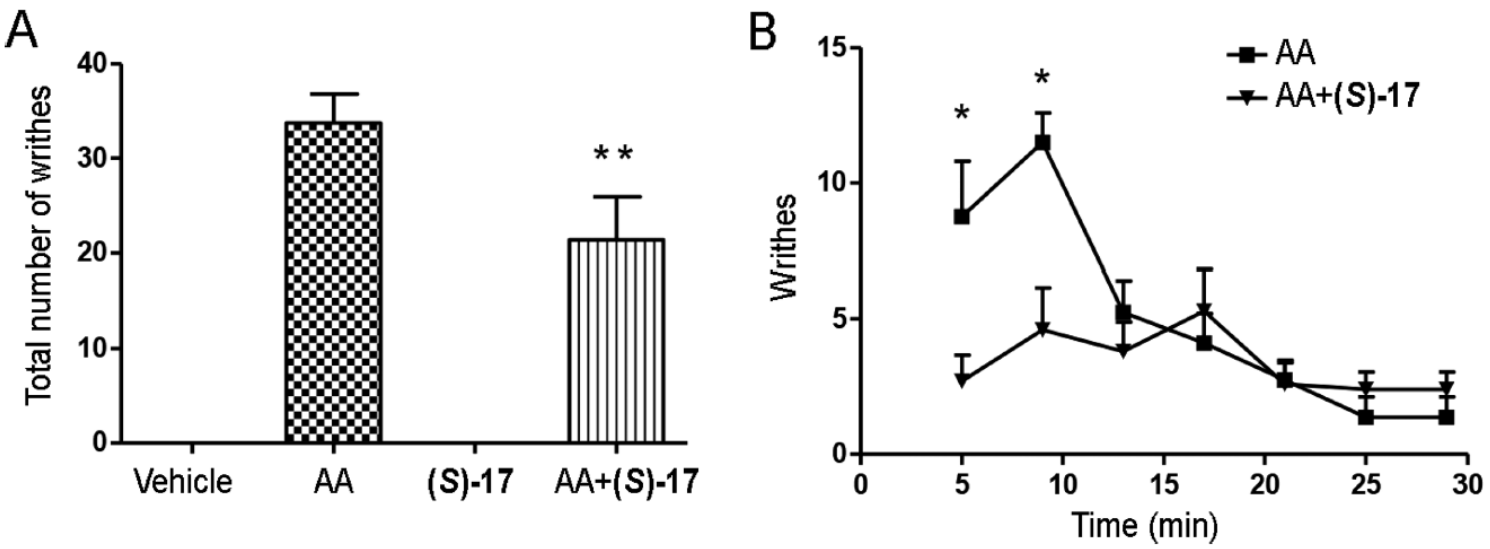

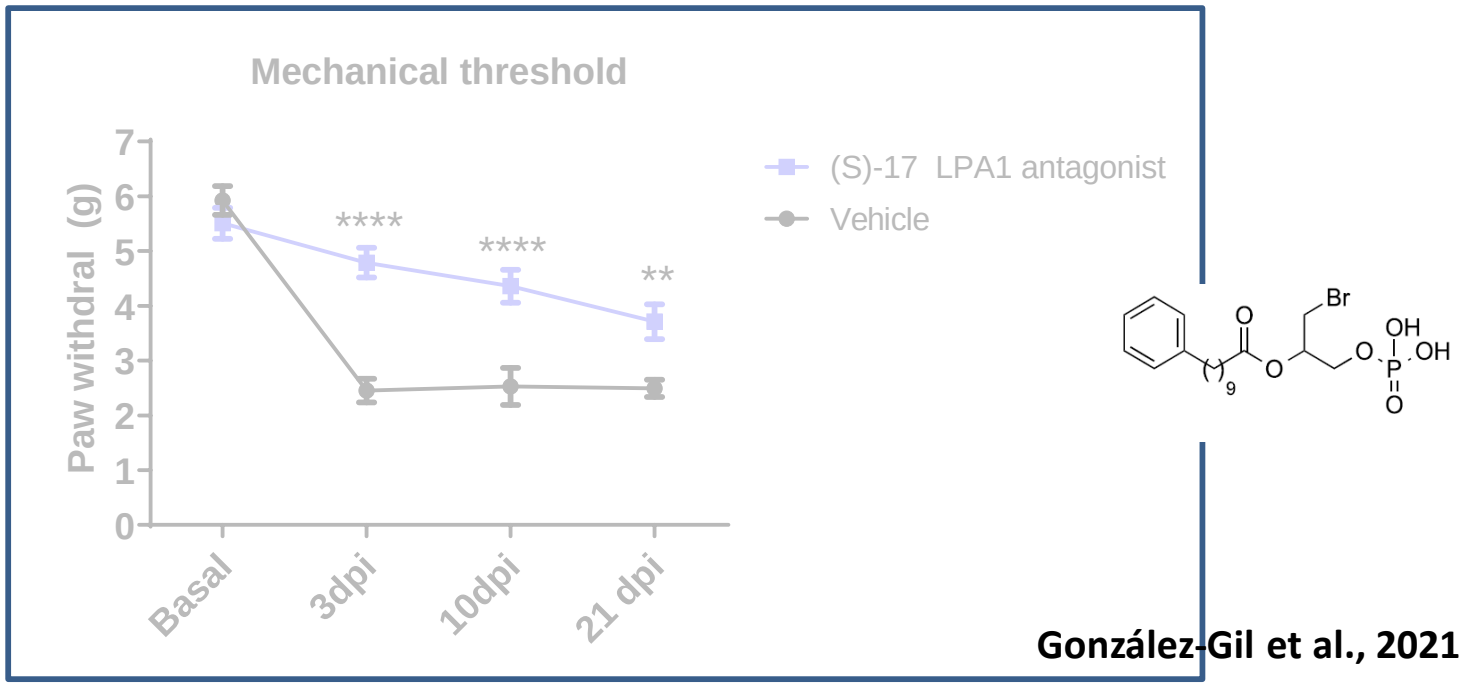

González-Gil et al., 2021

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MINISTERIO DE ECONOMIA Y COMPETITIVIDAD

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Enfermedades Neurodegenerativas

MARIE CURIE ACTIONS

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